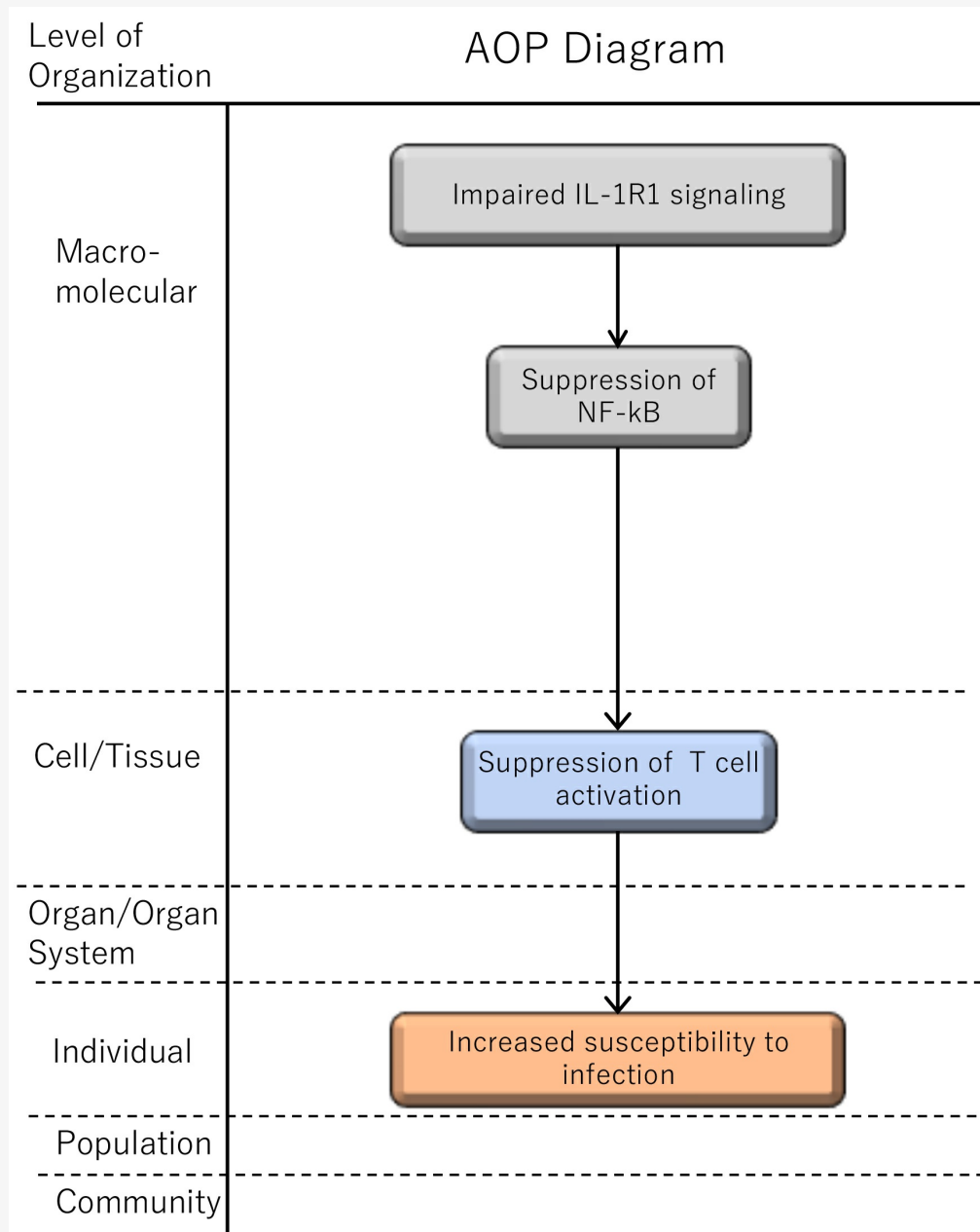


AOP ID and Title:

AOP 277: Impaired IL-1R1 signaling leading to increased susceptibility to infection

Short Title: IL-1 inhibition**Graphical Representation****Authors**

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Abstract

The pleiotropic cytokine IL-1 mediates its biological functions via association with the signaling receptor IL-1R1. These may include initiation of innate immunity as well as acquired immunity, which are essential for assistance of host defense against infection. The trimeric complex consists of IL-1, IL-1R1 and IL-1R3 (a coreceptor, formerly IL-1R accessory protein) allows for the approximation of the Toll-IL-1-Receptor (TIR) domains of each receptor chain. MyD88 then binds to the TIR domains. The binding of MyD88 triggers a cascade of kinases that produce a strong pro-inflammatory signal leading to activation of NF- κ B. The activation of NF- κ B plays a principal role in the immunological function of IL-1. Namely, it stimulates innate immunity such as activation of dendritic cells and macrophages. It also stimulates T cells via activated dendritic function or directly. The activation of T cells is crucial for B cell proliferation and their antibody production. The cooperation by T cells and B cells constitutes a main part of host defense against infection. Therefore, the impaired IL-1R1 signaling either by the decreased IL-1 production or the inhibition of IL-1 β binding to IL-1R1 by IL-1 receptor antagonist IL-1Ra or anti-IL-1 β antibody) results in the blockade of the effects of the pleiotropic cytokine IL-1 β leading to increased susceptibility to infection.

In this AOP, we selected the impaired IL-1R signaling as a molecular initiating event (MIE), and suppression of NF- κ B, suppression of T cell activation, and increased susceptibility to infection as key events (KE).

Background

The pleiotropic cytokine IL-1 mediates its biological functions via association with the signaling receptor IL-1R1. These may include initiation of innate immunity and assistance of host defense against infection, and sometimes, mediation of autoinflammatory, such as cryopyrin-associated periodic syndrome, neonatal-onset multisystem inflammatory disease and familial Mediterranean fever. The trimeric complex consists of IL-1, IL-1R1 and IL-1R3 (a coreceptor, formerly IL-1R accessory protein) allows for the approximation of the Toll-IL-1-Receptor (TIR) domains of each receptor chain. MyD88 then binds to the TIR domains. The binding of MyD88 triggers a cascade of kinases that produce a strong pro-inflammatory signal leading to activation of NF- κ B and fundamental inflammatory responses such as the induction of cyclooxygenase type 2, production of multiple cytokines and chemokines, increased expression of adhesion molecules, or synthesis of nitric oxide. (Dinarello, 2018; Weber et al., 2010a, b).

Molecules like nuclear or mitochondrial DNA, adenosine triphosphate (ATP), uridine triphosphate (UTP), uric acid and high mobility group box 1 (HMGB1) are classified as damage associated molecular patterns (DAMPs). DAMPs are secreted or produced upon cellular injury or death and induce sterile inflammation. On the other hand, bacterial products like lipopolysaccharide (LPS), peptidoglycans, lipoprotein flagellins, bacterial RNA and DNA are some of the well-characterized pathogen associated molecular patterns (PAMPs). These DAMPs and PAMPs with a few exceptions bind to pattern recognition receptors (PRRs) such as toll-like receptor (TLRs) and nucleotide oligomerization domain (NOD) like receptors (NLRs). Proinflammatory mediators such as DAMPs, PAMPs, and various inflammatory cytokines or mediators including IL-1 β itself activate innate immune mechanisms in the host leading to IL-1 β production (Handa et al., 2016; Newton and Dixit, 2012; Yang et al., 2017). Besides transcriptional regulation and posttranscriptional level by RNA-binding proteins, pro-IL-1 β protein requires proteolytic cleavage by active caspase-1 as the effector component of stimulation-induced multi-protein inflammasomes to acquire functional activity. Altogether, these different layers of regulation allow to fine tune IL-1 β production under different pathophysiological conditions (Bent et al., 2018).

Therefore, the inhibition of various targets in different layers from the stimulation of PRRs or the receptors of proinflammatory cytokines, e.g., IL-1, IL-18, or TNFa, to the activation of NF- κ B or the inhibition of posttranscriptional regulation of pro-IL-1 β cause impaired IL-1R1 signaling. In addition, since IL-1 also mediates autoinflammatory syndromes, such as cryopyrin-associated periodic syndrome, neonatal-onset multisystem inflammatory disease and familial Mediterranean fever, several inhibitors against IL-1R1 have been developed. They are IL-1 receptor antagonist IL-1Ra, anakinumab (anti-IL-1 β antibody) and rilonacept (soluble IL-1R). Several reports described that the administration of these drugs led to increased susceptibility to infection (De Benedetti et al., 2018; Fleischmann et al., 2003; Genovese et al., 2004; Imagawa et al., 2013; Kullenberg et al., 2016; Lachmann et al., 2009; Lequerre et al., 2008; Migkos et al., 2015; Schlesinger et al., 2012; Yokota et al., 2017). In addition to these human data, the experiments using knockout mice revealed that the lack of IL-1 signaling led to bacterial, tuberculosis or viral infection (Guler et al., 2011; Horino et al., 2009; Juffermans et al., 2000; Tian et al., 2017; Yamada et al., 2000).

Summary of the AOP

Events

Molecular Initiating Events (MIE), Key Events (KE), Adverse Outcomes (AO)

Sequence	Type	Event ID	Title	Short name
1	MIE	1700	Impaired IL-1R1 signaling	Impaired IL-1R1 signaling
2	KE	202	Inhibition, Nuclear factor kappa B (NF-κB)	Inhibition, Nuclear factor kappa B (NF- κ B)
3	KE	1702	Suppression of T cell activation	Suppression of T cell activation

Sequence	Type	Event ID	Title	Short name
1	1	1	Increase, Increased susceptibility to infection	Increase, Increased susceptibility to infection

Key Event Relationships

Upstream Event	Relationship Type	Downstream Event	Evidence	Quantitative Understanding
Impaired IL-1R1 signaling	adjacent	Inhibition, Nuclear factor kappa B (NF-kB)	High	Moderate
Inhibition, Nuclear factor kappa B (NF-kB)	adjacent	Suppression of T cell activation	High	Moderate
Suppression of T cell activation	adjacent	Increase, Increased susceptibility to infection	High	High

Stressors

Name	Evidence
IL-1 receptor antagonist IL-1Ra (Anakinra)	High
anti-IL-1b antibody (Canakinumab)	High
soluble IL-1R (Rilonacept)	High
anti-IL-1b antibody (Gevokizumab)	High
Dexamethasone	High
minocycline	High
Belnacasan (VX-765)	High
Pralnacasan (VX-740, HMR3480)	High
cinnamic aldehyde	High
Dimethyl fumarate	High
curcumin	High
iguratimod	High
(-)-Epigallocatechin gallate	High
TAK-242	High
IRAK4 inhibitors	High
Dehydroxymethylepoxyquinomicin (DHMEQ)	High

Overall Assessment of the AOP

Domain of Applicability

Life Stage Applicability

Life Stage	Evidence
Not Otherwise Specified	High

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
Homo sapiens	Homo sapiens	High	NCBI
Mus musculus	Mus musculus	High	NCBI
Rattus norvegicus	Rattus norvegicus	High	NCBI

Sex Applicability

Sex	Evidence
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Mixed Sex Evidence

Although sex differences in immune responses are well known (Klein and Flanagan, 2016), there is no reports regarding the sex difference in IL-1 production, IL-1 function or susceptibility to infection as adverse effect of IL-1 blocking agent. Again, age-dependent difference in IL-1 signaling is not known.

The IL1B gene is conserved in chimpanzee, Rhesus monkey, dog, cow, mouse, rat, and frog (<https://www.ncbi.nlm.nih.gov/homologene/481>), and the Myd88 gene is conserved in human, chimpanzee, Rhesus monkey, dog, cow, rat, chicken, zebrafish, mosquito, and frog (https://www.ncbi.nlm.nih.gov/homologene?Db=homologene&Cmd=Retrieve&list_uids=1849).

The NFKB1 gene is conserved in chimpanzee, Rhesus monkey, dog, cow, mouse, rat, chicken, and frog.

275 organisms have orthologs with human gene NFKB1.

(<https://www.ncbi.nlm.nih.gov/gene/4790>)

These data suggest that the proposed AOP regarding inhibition of IL-1 signaling is not dependent on life stage, sex, age or species.

Essentiality of the Key Events

The experiments using knockout mice revealed that the deficiency of IL-1 signaling led to bacterial, tuberculosis or viral infection (Guler et al., 2011; Horino et al., 2009; Juffermans et al., 2000; Tian, Jin and Dubin, 2017; Yamada et al., 2000).

IL-1 receptor antagonist IL-1Ra was purified in 1990, and the cDNA reported that same year. IL-1Ra binds IL-1R but does not initiate IL-1 signal transduction (Dripps et al., 1991). Recombinant IL-1Ra (generic anakinra) is fully active in blocking the IL-1R1, and therefore, the activities of IL-1 α and IL-1 β . Anakinra is approved for the treatment of rheumatoid arthritis and cryopyrin-associated periodic syndrome (CAPS). Since its introduction in 2002 for the treatment of rheumatoid arthritis, anakinra has had a remarkable record of safety. However, Fleischmann et al. (Fleischmann et al., 2003) reported that serious infectious episodes were observed more frequently in the anakinra group (2.1% versus 0.4% in the placebo group) and other authors reported the increased susceptibility to bacterial or tuberculosis infection (Genovese et al., 2004; Kullenberg et al., 2016; Lequerre et al., 2008; Migkos et al., 2015). As IL-1 signaling antagonists, two drugs went up to the market, canakinumab (anti-IL-1 β antibody) and rilonacept (soluble IL-1R). Several reports described that the administration of these drugs led to increased susceptibility to infection (De Benedetti et al., 2018; Imagawa et al., 2013; Lachmann et al., 2009; Schlesinger et al., 2012).

In a similar way, defect of MyD88 signaling caused by knockout of mice gene or deficiency in human patient leads to the increased susceptibility to bacterial or tuberculosis infection (von Bernuth et al., 2012).

Mice lacking NF- κ B p50 are unable effectively to clear *L. monocytogenes* and are more susceptible to infection with *S. pneumoniae* (Sha et al., 1995).

Weight of Evidence Summary

The recent review of IL-1 pathway by Weber et al. has clearly described the intracellular signaling event from the binding of IL-1 α or IL-1 β to IL-1R to the activation of NF- κ B through the assemble of MyD88 to the trimeric complex composed of IL-1, IL-1R1, and IL-1RacP. The sequentiality and essentiality of each signaling molecule have been demonstrated by mice lacking relevant molecules (Dinarello, 2018; Weber, Wasiliew and Kracht, 2010a, b).

There were several reports that described that administration of IL-1R antagonist or neutralizing antibody led to the suppression of downstream phenomena, which included internalization of IL-1 (Dripps et al., 1991), production of PGE₂ (Hannum et al., 1990; Seckinger et al., 1990), IL-6 (Goh et al., 2014), and T cell proliferation (Seckinger et al., 1990).

Several reports described that the administration of IL-1 receptor antagonist IL-1Ra, canakinumab (anti-IL-1 β antibody) and rilonacept (soluble IL-1R) led to increased susceptibility to infection (De Benedetti et al., 2018; Fleischmann et al., 2003; Genovese et al., 2004; Imagawa et al., 2013; Kullenberg et al., 2016; Lachmann et al., 2009; Lequerre et al., 2008; Migkos et al., 2015; Schlesinger et al., 2012; Yokota et al., 2017). In addition to these human data, the experiments using knockout mice revealed that the lack of IL-1 signaling led to bacterial, tuberculosis or viral infection (Guler et al., 2011; Horino et al., 2009; Juffermans et al., 2000; Tian, Jin and Dubin, 2017; Yamada et al., 2000). Moreover, polymorphism of IL-1 β or IL-1Ra leads to the increased susceptibility to tuberculosis, severe sepsis or fungal infection (Fang et al., 1999; Motsinger-Reif et al., 2010; Wojtowicz et al., 2015).

Biological plausibility

Inhibition of IL-1 binding to IL-1 receptor leads to Inhibition, Nuclear factor kappa B (NF- κ B)

IL-1 α and IL-1 β independently bind the type I IL-1 receptor (IL-1R1), which is ubiquitously expressed. The IL-1R3 (formerly IL-1R accessory protein (IL-1RacP)) serves as a co-receptor that is required for signal transduction of IL-1/IL-1R1 complexes.

The initial step in IL-1 signal transduction is a ligand-induced conformational change in the first extracellular domain of the IL-1RI that facilitates recruitment of IL-1R3. The trimeric complex rapidly assembles two intracellular signaling proteins, myeloid differentiation primary response gene 88 (MYD88) and interleukin-1 receptor-activated protein kinase (IRAK) 4. This is paralleled by the (auto)phosphorylation of IRAK4, which subsequently phosphorylates IRAK1 and IRAK2, and then this is followed by the recruitment and oligomerization of tumor necrosis factor-associated factor (TRAF) 6. Activation of NF- κ B by IL-1 requires the activation of inhibitor of nuclear factor B (I κ B) kinase 2 (IKK2). Activated IKK phosphorylates I κ B α , which promotes its K48-linked polyubiquitination and subsequent degradation by the proteasome. I κ B destruction allows the release of p50 and p65 NF- κ B subunits and their nuclear translocation, which is the central step in activation of NF- κ B. Both NF- κ Bs bind to a conserved DNA motif that is found in numerous IL-1-responsive genes. (Weber et al., 2010a, b)

Inhibition, Nuclear factor kappa B (NF- κ B) leads to Suppression of T cell activation

In T lineage cells, the temporal regulation of NF- κ B controls the stepwise differentiation and antigen-dependent selection of conventional and specialized subsets of T cells in response to T cell receptor and costimulatory, cytokines and growth factor signals. Cytokines include cytokines produced from macrophage or monocyte such as IL-1 β . (Gerondakis et al., 2014)

Suppression of T cell activation leads to Increase, Increased susceptibility to infection

First type immunity drives resistance to viruses and intracellular bacteria, such as *Listeria monocytogenes*, *Salmonella* spp. and *Mycobacteria* spp., as well as to intracellular protozoan parasites such as *Leishmania* spp. The T helper 1 signature cytokine interferon- γ has a central role in triggering cytotoxic mechanisms including macrophage polarization towards an antimicrobial response associated with the production of high levels of reactive oxygen species and reactive nitrogen species, activation of CD8 cytotoxic T lymphocytes and natural killer cells to kill infected cells via the perforin and/or granzyme B-dependent lytic pathway or via the ligation of surface death receptors; and B cell activation towards the production of cytolytic antibodies that target infected cells for complement and Fc receptor-mediated cellular cytotoxicity.

Resistance to extracellular metazoan parasites and other large parasites is mediated and/or involves second type immunity. Pathogen neutralization is achieved via different mechanisms controlled by T 2 signature cytokines, including interleukin-4, IL-5 and IL-13, and by additional type 2 cytokines such as thymic stromal lymphopoietin, IL-25 or IL-33, secreted by damaged cell. T 2 signature cytokines drive B cell activation towards the production of high-affinity pathogen-specific IgG1 and IgE antibodies that function via Fc-dependent mechanisms to trigger the activation of eosinophils, mast cells and basophils, expelling pathogens across epithelia.

T17 immunity confers resistance to extracellular bacteria such as *Klebsiella pneumoniae*, *Escherichia coli*, *Citrobacter rodentium*, *Bordetella pertussis*, *Porphyromonas gingivalis* and *Streptococcus pneumoniae*, and also to fungi such as *Candida albicans*, *Coccidioides posadasii*, *Histoplasma capsulatum* and *Blastomyces dermatitidis*. Activation of T 17 cells by cognate T cell receptor (TCR-MHC class II interactions and activation of group 3 innate lymphoid cells (ILC3s) via engagement of IL-1 receptor (IL-1R) by IL-1 β secreted from damaged cells lead to the recruitment and activation of neutrophils. T 17 immunopathology is driven to a large extent by products of neutrophil activation, such as ROS and elastase (reviewed by Soares et al. (Soares et al., 2017).

Based on these evidences, the insufficient T cell or B cell function causes impaired resistance to infection.

Empirical support

1. Impaired IL-1R signaling.

Decreased production of IL-1 or inhibition of the binding of IL-1 to IL-1R impair IL-1R signaling.

1-1. Decreased IL-1 production

Decreased IL-1 production by macrophages or dendritic cells can be induced by suppressed IL-1 β mRNA induction or suppressed maturation of pro-IL-1 β . Dexamethasone is one of the representative drugs that significantly suppress IL-1 β production from monocytes (Finch-Arietta and Cochran, 1991). Other than dexamethasone, the inhibition of various targets in different layers from the stimulation of PRRs or the receptors of proinflammatory cytokines to the activation of NF- κ B or the inhibition of posttranscriptional regulation of pro-IL-1 β cause impaired decreased IL-1 β production.

Quite a few compounds have been reported to inhibit NF- κ B signaling by several different mechanisms reviewed by Fuchs (Fuchs, 2010). The list of representative chemicals and their mechanism to inhibit NF- κ B is shown in Table 1. In fact, dimethyl fumarate inhibits the activation of NF- κ B, resulting in a loss of proinflammatory cytokine production, distorted maturation and function of antigen-presenting cells, and immune deviation of T helper cells (Th) from the type 1 (Th1) and type 17 (Th17) profiles to a type 2 (Th2) phenotype (McGuire et al., 2016; Peng et al., 2012). Several studies have shown intriguing pharmacologic effects associated with curcumin, which inhibits NF- κ B expression by regulating NF- κ B/I κ B pathway and down-regulates expression of pro-inflammatory cytokines, such as IL-1, IL-6, IL-8, and TNF α (Wang et al., 2018). Igaratimod, a methanesulfonanilide, that is a novel disease-modifying antirheumatic drug, inhibits NF- κ B but not its inhibitor, I κ B α , and inhibits the production of IL-1 β (Mucke, 2012). Epigallocatechin gallate (EGCG) has been reported to inhibit NF- κ B activation through inhibition of p65 phosphorylation (Wheeler et al., 2004) and suppress the production of LPS-stimulated IL-1 β (Wang et al., 2020). DHMEQ inhibits LPS-induced NF- κ B activation by inhibiting its nuclear translocation from the cytoplasm. It also inhibits LPS-induced secretion of IL-1 β (Suzuki and Umezawa, 2006).

Other than the inhibitors for NF- κ B signaling, which can be stimulated by various stimulations other than TLR4 stimulation, there are signaling molecules that are specific to TLR4 signaling, such as TLR4, Mal, TRAM, Myd88, IRAK4, and IRAK1/2 (Vallabhapurapu

and Karin, 2009). There are several chemicals that target some of these molecules, an inhibitors of TLR4 such as TAK-242 (Matsunaga et al., 2011) and various IRAK4 inhibitors (Lee et al., 2017). IRAK4 has recently attracted attention as a therapeutic target for inflammation and tumor diseases (Chaudhary, Robinson and Romero, 2015).

Beside transcriptional regulation of IL-1 β production, minocycline, and two prodrugs, pralnacasan (VX-740) and belnacasan (VX-765) that are orally absorbed and converted into the active principle, VRT-018858 and VRT-043198, respectively (Fenini, Contassot and French, 2017) suppress IL-1 signaling by the inhibition of caspase-1 activation. Caspase-1 is an essential enzyme for maturation of pro- IL-1 β and the secretion of mature IL-1 β (Vincent and Mohr, 2007). Recently, it has been reported that cinnamic aldehyde suppresses serum IL-1 β level in endotoxin poisoning mice (Xu et al., 2017).

1-2. Blocking of binding of IL-1 to IL-1R1

IL-1 α and IL-1 β independently bind the type I IL-1 receptor (IL-1R1), which is ubiquitously expressed. IL-1Ra binds IL-1R but does not initiate IL-1 signal transduction (Dripps et al., 1991). Recombinant IL-1Ra (anakinra) is fully active in blocking the IL-1R1, and therefore, the biological activities of IL-1 α and IL-1 β . The binding of IL-1 α and IL-1 β to IL-1R1 can be suppressed by soluble IL-1R like rilonacept (Kapur and Bonk, 2009). The binding of IL-1 β to IL-1R1 can be inhibited by anti-IL-1 β antibody (canakinumab and gevokizumab) (Church and McDermott, 2009) (Roell et al., 2010).

Several reports described that the administration of IL-1 receptor antagonist IL-1Ra, canakinumab (anti-IL-1 β antibody) and rilonacept (soluble IL-1R) led to increased susceptibility to infection (De Benedetti et al., 2018; Fleischmann et al., 2003; Genovese et al., 2004; Imagawa et al., 2013; Kullenberg et al., 2016; Lachmann et al., 2009; Lequerre et al., 2008; Migkos et al., 2015; Schlesinger et al., 2012; Yokota et al., 2017).

1-3. IL-1 deficient mice

In addition to these human data, the experiments using knockout mice revealed that the lack of IL-1 signaling led to bacterial, tuberculosis or viral infection (Guler et al., 2011; Horino et al., 2009; Juffermans et al., 2000; Tian, Jin and Dubin, 2017; Yamada et al., 2000). Moreover, polymorphism of IL-1 β or IL-1Ra leads to the increased susceptibility to tuberculosis, severe sepsis or fungal infection (Fang et al., 1999; Motsinger-Reif et al., 2010; Wojtowicz et al., 2015). RelB deficient mice had an impaired cellular immunity, as observed in contact sensitivity reaction (Weih et al., 1995).

Quantitative Consideration

IL-1Ra blocks IL-1 signaling:

IL-1Ra alone at concentrations as high as 1 mg/mL did not induce IL-1 α , IL-1 β , TNF α , or IL-6 synthesis. Suppression of IL-1-induced IL-1, TNF α , or IL-6 synthesis was dose-dependent ($P \leq .0001$). At a twofold molar excess, IL-1Ra inhibited IL-1-induced IL-1 or TNF α synthesis by 50% ($P < .01$); an equimolar concentration of IL-1Ra inhibited synthesis of these two cytokines by over 20% ($P < .05$). A 10-fold molar excess of IL-1Ra over IL-1 β reduced IL-1 β -induced IL-1 α by 95% ($P = .01$) and IL-1 α -induced IL-1 β by 73% ($P < .01$). In elutriated monocytes, a 10-fold molar excess of IL-1Ra reduced IL-1 β -induced IL-1 α by 82% ($P < .05$), TNF α by 64% ($P = .05$), and IL-6 by 47% ($P < .05$). (Granowitz et al., 1992)

Canakinumab (ACZ885, Ilaris):

The antibody binds to human IL-1 β with high affinity (about 40 pmol/l). The antibody was found to neutralize the bioactivity of human IL-1 β on primary human fibroblasts in vitro 44.6 pmol/l (7.1 ± 0.56 ng/ml; $n = 6$) of ED50. Application of Canakinumab intraperitoneally 2 hours before injecting the IL-1 β producing cells completely suppressed joint swelling (0.06 mg/kg of EC50) (Alten et al., 2008).

Primary human fibroblasts are stimulated with recombinant IL-1b or conditioned medium obtained from LPS-stimulated human PBMCs in the presence of various concentrations of Cankinumab or IL-1RA ranging from 6 to 18,000 pM. Supernatant is taken after 16 h stimulation and assayed for IL-6 by ELISA. Canakinumab typically have 1 nM or less of EC50 for inhibition of IL-6 production (Canakinumab Patent Application WO02/16436.)

Rilonacept (IL-1 Trap, Arcalyst):

Incubation of the human MRC5 fibroblastic cell line with IL-1 β induces secretion of IL-6. At a constant amount of IL-1 β (4 pM), the IC50 of the IL-1 trap is ~2 pM. Another unique property of the IL-1 trap is that it not only blocks IL-1 β , but also blocks IL-1 α with high affinity ($KD = \sim 3$ pM; data not shown). The titration curve of IL-1 trap in the presence of 10 pM IL-1 β shows an IC50 of 6.5 pM, which corresponds to a calculated KD of 1.5 pM (This affinity is 100 times higher than that of the soluble single component receptor IL-1RI (Economides et al., 2003).

Considerations for Potential Applications of the AOP (optional)

The impaired IL-1 signaling can lead to decreased host resistance to various infections. Therefore, the test guideline to detect chemicals that decrease IL-1 signaling is required to support regulatory decision-making. This AOP can promote the understanding of the usefulness of the test guideline.

References

- Alten, R., Gram, H., Joosten, L.A., et al. (2008), The human anti-IL-1 beta monoclonal antibody ACZ885 is effective in joint inflammation models in mice and in a proof-of-concept study in patients with rheumatoid arthritis. *Arthritis Res Ther* 10: R67, 10.1186/ar2438
- Bent, R., Moll, L., Grabbe, S., et al. (2018), Interleukin-1 Beta-A Friend or Foe in Malignancies? *Int J Mol Sci* 19, 10.3390/ijms19082155
- Chaudhary, D., Robinson, S., Romero, D.L. (2015), Recent advances in the discovery of small molecule inhibitors of interleukin-1 receptor-associated kinase 4 (IRAK4) as a therapeutic target for inflammation and oncology disorders. *J Med Chem* 58: 96-110, 10.1021/jm5016044
- Church, L.D., McDermott, M.F. (2009), Canakinumab, a fully-human mAb against IL-1beta for the potential treatment of inflammatory disorders. *Curr Opin Mol Ther* 11: 81-89,
- De Benedetti, F., Gattorno, M., Anton, J., et al. (2018), Canakinumab for the Treatment of Autoinflammatory Recurrent Fever Syndromes. *N Engl J Med* 378: 1908-1919, 10.1056/NEJMoa1706314
- Dinarello, C.A. (2018), Overview of the IL-1 family in innate inflammation and acquired immunity. *Immunol Rev* 281: 8-27, 10.1111/imr.12621
- Dripps, D.J., Brandhuber, B.J., Thompson, R.C., et al. (1991), Interleukin-1 (IL-1) receptor antagonist binds to the 80-kDa IL-1 receptor but does not initiate IL-1 signal transduction. *J Biol Chem* 266: 10331-10336,
- Economides, A.N., Carpenter, L.R., Rudge, J.S., et al. (2003), Cytokine traps: multi-component, high-affinity blockers of cytokine action. *Nat Med* 9: 47-52, 10.1038/nm811
- Fang, X.M., Schroder, S., Hoefft, A., et al. (1999), Comparison of two polymorphisms of the interleukin-1 gene family: interleukin-1 receptor antagonist polymorphism contributes to susceptibility to severe sepsis. *Crit Care Med* 27: 1330-1334, 10.1097/00003246-199907000-00024
- Fenini, G., Contassot, E., French, L.E. (2017), Potential of IL-1, IL-18 and Inflammasome Inhibition for the Treatment of Inflammatory Skin Diseases. *Front Pharmacol* 8: 278, 10.3389/fphar.2017.00278
- Finch-Arietta, M.B., Cochran, F.R. (1991), Cytokine production in whole blood ex vivo. *Agents Actions* 34: 49-52,
- Fleischmann, R.M., Schechtman, J., Bennett, R., et al. (2003), Anakinra, a recombinant human interleukin-1 receptor antagonist (r-metHuIL-1ra), in patients with rheumatoid arthritis: A large, international, multicenter, placebo-controlled trial. *Arthritis Rheum* 48: 927-934, 10.1002/art.10870
- Fuchs, O. (2010), Transcription factor NF-kappaB inhibitors as single therapeutic agents or in combination with classical chemotherapeutic agents for the treatment of hematologic malignancies. *Curr Mol Pharmacol* 3: 98-122, 10.2174/1874467211003030098
- Genovese, M.C., Cohen, S., Moreland, L., et al. (2004), Combination therapy with etanercept and anakinra in the treatment of patients with rheumatoid arthritis who have been treated unsuccessfully with methotrexate. *Arthritis Rheum* 50: 1412-1419, 10.1002/art.20221
- Gerondakis, S., Fulford, T.S., Messina, N.L., et al. (2014), NF-kappaB control of T cell development. *Nat Immunol* 15: 15-25, 10.1038/ni.2785
- Goh, A.X., Bertin-Maghit, S., Ping Yeo, S., et al. (2014), A novel human anti-interleukin-1beta neutralizing monoclonal antibody showing in vivo efficacy. *MAbs* 6: 765-773, 10.4161/mabs.28614
- Granowitz, E.V., Clark, B.D., Vannier, E., et al. (1992), Effect of interleukin-1 (IL-1) blockade on cytokine synthesis: I. IL-1 receptor antagonist inhibits IL-1-induced cytokine synthesis and blocks the binding of IL-1 to its type II receptor on human monocytes. *Blood* 79: 2356-2363,
- Guler, R., Parihar, S.P., Spohn, G., et al. (2011), Blocking IL-1alpha but not IL-1beta increases susceptibility to chronic Mycobacterium tuberculosis infection in mice. *Vaccine* 29: 1339-1346, 10.1016/j.vaccine.2010.10.045
- Handa, P., Vemulakonda, A., Kowdley, K.V., et al. (2016), Mitochondrial DNA from hepatocytes as a ligand for TLR9: Drivers of nonalcoholic steatohepatitis? *World J Gastroenterol* 22: 6965-6971, 10.3748/wjg.v22.i31.6965
- Hannum, C.H., Wilcox, C.J., Arend, W.P., et al. (1990), Interleukin-1 receptor antagonist activity of a human interleukin-1 inhibitor. *Nature* 343: 336-340, 10.1038/343336a0
- Ho, S.C., Chang, Y.H., Chang, K.S. (2018), Structural Moieties Required for Cinnamaldehyde-Related Compounds to Inhibit Canonical IL-1beta Secretion. *Molecules* 23, 10.3390/molecules23123241
- Horino, T., Matsumoto, T., Ishikawa, H., et al. (2009), Interleukin-1 deficiency in combination with macrophage depletion increases

- susceptibility to *Pseudomonas aeruginosa* bacteremia. *Microbiol Immunol* 53: 502-511, 10.1111/j.1348-0421.2009.00143.x
- Huang, H., Wang, Y. (2017), The protective effect of cinnamaldehyde on lipopolysaccharide induced acute lung injury in mice. *Cell Mol Biol (Noisy-le-grand)* 63: 58-63, 10.14715/cmb/2017.63.8.13
- Imagawa, T., Nishikomori, R., Takada, H., et al. (2013), Safety and efficacy of canakinumab in Japanese patients with phenotypes of cryopyrin-associated periodic syndrome as established in the first open-label, phase-3 pivotal study (24-week results). *Clin Exp Rheumatol* 31: 302-309,
- Jeon, Y.J., Han, S.H., Lee, Y.W., et al. (2000), Dexamethasone inhibits IL-1 beta gene expression in LPS-stimulated RAW 264.7 cells by blocking NF-kappa B/Rel and AP-1 activation. *Immunopharmacology* 48: 173-183,
- Juffermans, N.P., Florquin, S., Camoglio, L., et al. (2000), Interleukin-1 signaling is essential for host defense during murine pulmonary tuberculosis. *J Infect Dis* 182: 902-908, 10.1086/315771
- Kapur, S., Bonk, M.E. (2009), Rilonacept (arcalyst), an interleukin-1 trap for the treatment of cryopyrin-associated periodic syndromes. *P t* 34: 138-141,
- Klein, S.L., Flanagan, K.L. (2016), Sex differences in immune responses. *Nat Rev Immunol* 16: 626-638, 10.1038/nri.2016.90
- Kullenberg, T., Lofqvist, M., Leinonen, M., et al. (2016), Long-term safety profile of anakinra in patients with severe cryopyrin-associated periodic syndromes. *Rheumatology (Oxford)* 55: 1499-1506, 10.1093/rheumatology/kew208
- Lachmann, H.J., Kone-Paut, I., Kuemmerle-Deschner, J.B., et al. (2009), Use of canakinumab in the cryopyrin-associated periodic syndrome. *N Engl J Med* 360: 2416-2425, 10.1056/NEJMoa0810787
- Lee, K.L., Ambler, C.M., Anderson, D.R., et al. (2017), Discovery of Clinical Candidate 1-[(2S,3S,4S)-3-Ethyl-4-fluoro-5-oxopyrrolidin-2-yl]methoxy-7-methoxyisoquinoline-6-carboxamide (PF-06650833), a Potent, Selective Inhibitor of Interleukin-1 Receptor Associated Kinase 4 (IRAK4), by Fragment-Based Drug Design. *J Med Chem* 60: 5521-5542, 10.1021/acs.jmedchem.7b00231
- Lequerre, T., Quartier, P., Rosellini, D., et al. (2008), Interleukin-1 receptor antagonist (anakinra) treatment in patients with systemic-onset juvenile idiopathic arthritis or adult onset Still disease: preliminary experience in France. *Ann Rheum Dis* 67: 302-308, 10.1136/ard.2007.076034
- Matsunaga, N., Tsuchimori, N., Matsumoto, T., et al. (2011), TAK-242 (resatorvid), a small-molecule inhibitor of Toll-like receptor (TLR) 4 signaling, binds selectively to TLR4 and interferes with interactions between TLR4 and its adaptor molecules. *Mol Pharmacol* 79: 34-41, 10.1124/mol.110.068064
- McGuire, V.A., Ruiz-Zorrilla Diez, T., Emmerich, C.H., et al. (2016), Dimethyl fumarate blocks pro-inflammatory cytokine production via inhibition of TLR induced M1 and K63 ubiquitin chain formation. *Sci Rep* 6: 31159, 10.1038/srep31159
- Migkos, M.P., Somarakis, G.A., Markatseli, T.E., et al. (2015), Tuberculous pyomyositis in a rheumatoid arthritis patient treated with anakinra. *Clin Exp Rheumatol* 33: 734-736,
- Morand, E.F., Rickard, D., Goulding, N.J. (1993), Lack of involvement of lipocortin 1 in dexamethasone suppression of IL-1 release. *Mediators Inflamm* 2: 49-52, 10.1155/s0962935193000067
- Motsinger-Reif, A.A., Antas, P.R., Oki, N.O., et al. (2010), Polymorphisms in IL-1beta, vitamin D receptor Fok1, and Toll-like receptor 2 are associated with extrapulmonary tuberculosis. *BMC Med Genet* 11: 37, 10.1186/1471-2350-11-37
- Mucke, H.A. (2012), Igaratimod: a new disease-modifying antirheumatic drug. *Drugs Today (Barc)* 48: 577-586, 10.1358/dot.2012.48.9.1855758
- Newton, K., Dixit, V.M. (2012), Signaling in innate immunity and inflammation. *Cold Spring Harb Perspect Biol* 4, 10.1101/cshperspect.a006049
- Peng, H., Guerau-de-Arellano, M., Mehta, V.B., et al. (2012), Dimethyl fumarate inhibits dendritic cell maturation via nuclear factor kappaB (NF-kappaB) and extracellular signal-regulated kinase 1 and 2 (ERK1/2) and mitogen stress-activated kinase 1 (MSK1) signaling. *J Biol Chem* 287: 28017-28026, 10.1074/jbc.M112.383380
- Qiu, H.B., Pan, J.Q., Zhao, Y.Q., et al. (1997), Effects of dexamethasone and ibuprofen on LPS-induced gene expression of TNF alpha, IL-1 beta, and MIP-1 alpha in rat lung. *Zhongguo Yao Li Xue Bao* 18: 165-168,
- Quartier, P. (2011), Interleukin-1 antagonists in the treatment of autoinflammatory syndromes, including cryopyrin-associated periodic syndrome. *Open Access Rheumatol* 3: 9-18, 10.2147/oarr.S6696
- Ravizza, T., Lucas, S.M., Balosso, S., et al. (2006), Inactivation of caspase-1 in rodent brain: a novel anticonvulsive strategy. *Epilepsia* 47: 1160-1168, 10.1111/j.1528-1167.2006.00590.x
- Roell, M.K., Issafras, H., Bauer, R.J., et al. (2010), Kinetic approach to pathway attenuation using XOMA 052, a regulatory therapeutic antibody that modulates interleukin-1beta activity. *J Biol Chem* 285: 20607-20614, 10.1074/jbc.M110.115790

- Schlesinger, N., Alten, R.E., Bardin, T., et al. (2012), Canakinumab for acute gouty arthritis in patients with limited treatment options: results from two randomised, multicentre, active-controlled, double-blind trials and their initial extensions. *Ann Rheum Dis* 71: 1839-1848, 10.1136/annrheumdis-2011-200908
- Seckinger, P., Kaufmann, M.T., Dayer, J.M. (1990), An interleukin 1 inhibitor affects both cell-associated interleukin 1-induced T cell proliferation and PGE2/collagenase production by human dermal fibroblasts and synovial cells. *Immunobiology* 180: 316-327, 10.1016/s0171-2985(11)80295-0
- Sha, W.C., Liou, H.C., Tuomanen, E.I., et al. (1995), Targeted disruption of the p50 subunit of NF-kappa B leads to multifocal defects in immune responses. *Cell* 80: 321-330,
- Soares, M.P., Teixeira, L., Moita, L.F. (2017), Disease tolerance and immunity in host protection against infection. *Nat Rev Immunol* 17: 83-96, 10.1038/nri.2016.136
- Stack, J.H., Beaumont, K., Larsen, P.D., et al. (2005), IL-converting enzyme/caspase-1 inhibitor VX-765 blocks the hypersensitive response to an inflammatory stimulus in monocytes from familial cold autoinflammatory syndrome patients. *J Immunol* 175: 2630-2634,
- Suzuki, E., Umezawa, K. (2006), Inhibition of macrophage activation and phagocytosis by a novel NF-kappaB inhibitor, dehydroxymethylepoxyquinomicin. *Biomed Pharmacother* 60: 578-586, 10.1016/j.biopha.2006.07.089
- Tian, T., Jin, M.Q., Dubin, K. (2017), IL-1R Type 1-Deficient Mice Demonstrate an Impaired Host Immune Response against Cutaneous Vaccinia Virus Infection. 198: 4341-4351, 10.4049/jimmunol.1500106
- Vallabhapurapu, S., Karin, M. (2009), Regulation and function of NF-kappaB transcription factors in the immune system. *Annu Rev Immunol* 27: 693-733, 10.1146/annurev.immunol.021908.132641
- van Furth, A.M., Seijmonsbergen, E.M., Langermans, J.A., et al. (1995), Effect of xanthine derivatives and dexamethasone on Streptococcus pneumoniae-stimulated production of tumor necrosis factor alpha, interleukin-1 beta (IL-1 beta), and IL-10 by human leukocytes. *Clin Diagn Lab Immunol* 2: 689-692,
- Vincent, J.A., Mohr, S. (2007), Inhibition of caspase-1/interleukin-1beta signaling prevents degeneration of retinal capillaries in diabetes and galactosemia. *Diabetes* 56: 224-230, 10.2337/db06-0427
- Wang, A.L., Yu, A.C., Lau, L.T., et al. (2005), Minocycline inhibits LPS-induced retinal microglia activation. *Neurochem Int* 47: 152-158, 10.1016/j.neuint.2005.04.018
- Wang, F., Han, Y., Xi, S., et al. (2020), Catechins reduce inflammation in lipopolysaccharide-stimulated dental pulp cells by inhibiting activation of the NF-kappaB pathway. *Oral Dis* 26: 815-821, 10.1111/odi.13290
- Wang, Y., Tang, Q., Duan, P., et al. (2018), Curcumin as a therapeutic agent for blocking NF-kappaB activation in ulcerative colitis. *Immunopharmacol Immunotoxicol* 40: 476-482, 10.1080/08923973.2018.1469145
- Weber, A., Wasiliew, P., Kracht, M. (2010a), Interleukin-1 (IL-1) pathway. *Sci Signal* 3: cm1, 10.1126/scisignal.3105cm1
- Weber, A., Wasiliew, P., Kracht, M. (2010b), Interleukin-1beta (IL-1beta) processing pathway. *Sci Signal* 3: cm2, 10.1126/scisignal.3105cm2
- Weih, F., Carrasco, D., Durham, S.K., et al. (1995), Multiorgan inflammation and hematopoietic abnormalities in mice with a targeted disruption of RelB, a member of the NF-kappa B/Rel family. *Cell* 80: 331-340,
- Weinberg, J.B., Mason, S.N., Wortham, T.S. (1992), Inhibition of tumor necrosis factor-alpha (TNF-alpha) and interleukin-1 beta (IL-1 beta) messenger RNA (mRNA) expression in HL-60 leukemia cells by pentoxifylline and dexamethasone: dissociation of acivicin-induced TNF-alpha and IL-1 beta mRNA expression from acivicin-induced monocytoid differentiation. *Blood* 79: 3337-3343,
- Wheeler, D.S., Catravas, J.D., Odoms, K., et al. (2004), Epigallocatechin-3-gallate, a green tea-derived polyphenol, inhibits IL-1 beta-dependent proinflammatory signal transduction in cultured respiratory epithelial cells. *J Nutr* 134: 1039-1044, 10.1093/jn/134.5.1039
- Wojtowicz, A., Gresnigt, M.S., Lecompte, T., et al. (2015), IL1B and DEFB1 Polymorphisms Increase Susceptibility to Invasive Mold Infection After Solid-Organ Transplantation. *J Infect Dis* 211: 1646-1657, 10.1093/infdis/jiu636
- Xu, F., Wang, F., Wen, T., et al. (2017), Inhibition of NLRP3 inflammasome: a new protective mechanism of cinnamaldehyde in endotoxin poisoning of mice. *Immunopharmacol Immunotoxicol* 39: 296-304, 10.1080/08923973.2017.1355377
- Yamada, H., Mizumo, S., Horai, R., et al. (2000), Protective role of interleukin-1 in mycobacterial infection in IL-1 alpha/beta double-knockout mice. *Lab Invest* 80: 759-767,
- Yang, Han, Z., Oppenheim, J.J. (2017), Alarmins and immunity. *Immunol Rev* 280: 41-56, 10.1111/imr.12577
- Yokota, S., Imagawa, T., Nishikomori, R., et al. (2017), Long-term safety and efficacy of canakinumab in cryopyrin-associated periodic syndrome: results from an open-label, phase III pivotal study in Japanese patients. *Clin Exp Rheumatol* 35 Suppl 108: 19-26,

Appendix 1

List of MIEs in this AOP

[Event: 1700: Impaired IL-1R1 signaling](#)

Short Name: Impaired IL-1R1 signaling

AOPs Including This Key Event

AOP ID and Name	Event Type
Aop:277 - Impaired IL-1R1 signaling leading to increased susceptibility to infection	MolecularInitiatingEvent

Stressors

Name
IL-1 receptor antagonist IL-1Ra (Anakinra)
anti-IL-1b antibody (Canakinumab)
soluble IL-1R (Rilonacept)
curcumin
iguratimod
epigallocatechin gallate
TAK-242
IRAK4 inhibitors
Dehydroxymethylepoxyquinomicin (DHMEQ)
Dimethyl fumarate
anti-IL-1b antibody (Gevokizumab)

Biological Context

Level of Biological Organization

Molecular

Cell term

Cell term

macrophage

Organ term

Organ term

immune system

Evidence for Perturbation by Stressor

Overview for Molecular Initiating Event

Dex inhibits IL-1 β gene expression in LPS-stimulated RAW 264.7 cells by blocking NF- κ B/Rel and AP-1 activation (Jeon et al., 2000).

Dex suppresses LPS-induced gene expression of IL-1 β in rat lung. (in vivo) (Qiu et al., 1997)

Dex inhibits the release of IL-1 β by human leukocyte stimulated with *Streptococcus pneumoniae* stimulation (van Furth et al., 1995).

Treatment of peripheral blood monocytes with 2 mg/ml LPS potently increased IL-1 β release ($p=0.001$) and Dex (10^{-7} M) significantly reduced both resting and stimulated IL-1 β release ($p=0.009$). (Morand et al., 1993)

Dex effectively blocks the glutamine antagonist acivicin-induced expression of IL-1 β mRNA by HL-60 leukemia cells (Weinberg et al., 1992)

Various inhibitors for NF- κ B, such as dimethyl fumarate, curcumin, iguratimod, epigallocatechin gallate (EGCG), DHMEQ inhibits LPS-induced NF- κ B activation and LPS-induced secretion of IL-1 β (McGuire et al., 2016; Peng et al., 2012) (Wang et al., 2018) (Mucke, 2012) (Wheeler et al., 2004) (Wang et al., 2020) (Suzuki and Umezawa, 2006) (Suzuki and Umezawa, 2006).

Several chemicals that target some of these molecules, an inhibitors of TLR4 such as TAK-242 (Matsunaga et al., 2011) and various IRAK4 inhibitors (Lee et al., 2017). IRAK4 has recently attracted attention as a therapeutic target for inflammation and tumor diseases.

LPS treatment induced a significant upregulation of the mRNA and release of IL-1 β from retinal microglia. Minocycline inhibited its releases. Thus, minocycline might exert its antiinflammatory effect on microglia by inhibiting the expression and release of IL-1 β (Wang et al., 2005).

Caspase-1 inhibition reduced the release of IL-1 β in organotypic slices exposed to LPS+ATP. Administration of pralnacasan (intracerebroventricular, 50 μ g) or belnacasan (intraperitoneal, 25–200 mg/kg) to rats blocked seizure-induced production of IL-1 β in the hippocampus, and resulted in a twofold delay in seizure onset and 50% reduction in seizure duration (Ravizza et al., 2006).

Belnacasan, an orally active IL-1 β converting enzyme/caspase-1 inhibitor, blocked IL-1 β secretion with equal potency in LPS-stimulated cells from familial cold urticarial associated syndrome and control subjects (Stack et al., 2005).

In LPS-induced acute lung injury (ALI) mice model, LPS induced inflammatory cytokines such as TNF- α , IL-6, IL-13 and IL-1 β were significantly decreased by cinnamaldehyde (CA) (Huang and Wang, 2017).

The suppressing capacities of six cinnamaldehyde-related compounds were evaluated and compared by using the LPS-primed and ATP-activated macrophages. At concentrations of 25–100 mM, cinnamaldehyde and 2-methoxy cinnamaldehyde dose-dependently inhibited IL-1 β secretion (Ho et al., 2018).

In vitro, CA decreased the levels of pro-IL-1 β and IL-1 β in cell culture supernatants, as well as the expression of NLRP3 and IL-1 β mRNA in cells. In vivo, CA decreased IL-1 β production in serum. Furthermore, CA suppressed LPS-induced NLRP3, p20, Pro-IL-1 β , P2X7 receptor (P2X7R) and cathepsin B protein expression in lung, as well as the expression of NLRP3 and IL-1 β mRNA (Xu et al., 2017).

IL-1Ra binds IL-1R but does not initiate IL-1 signal transduction (Dripps et al., 1991). Recombinant IL-1Ra (anakinra) is fully active in blocking the IL-1R1, and therefore, the biological activities of IL-1 α and IL-1 β . The binding of IL-1 α and IL-1 β to IL-1R1 can be suppressed by soluble IL-1R like rilonacept (Kapur and Bonk, 2009). The binding of IL-1 β to IL-1R1 can be inhibited by anti-IL-1 β antibody (canakinumab and gevokizumab) (Church and McDermott, 2009) (Roell et al., 2010).

IL-1 is known to mediate autoinflammatory syndrome, such as cryopyrin-associated periodic syndrome, neonatal-onset multisystem inflammatory disease and familial Mediterranean fever. Blocking of binding of IL-1 to IL-1R1 by anakinra, canakinumab, and rilonacept have been already used to treat these autoinflammatory syndrome associated with overactivation of IL-1 signaling (Quartier, 2011).

Domain of Applicability

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
Homo sapiens	Homo sapiens	High	NCBI
Mus musculus	Mus musculus	High	NCBI
Rattus norvegicus	Rattus norvegicus	High	NCBI

Life Stage Applicability

Life Stage	Evidence
All life stages	High

Sex Applicability

Sex	Evidence
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Unspecific High Evidence

Although sex differences in immune responses are well known (Klein and Flanagan, 2016), there is no reports regarding the sex difference in IL-1 production, IL-1 function or susceptibility to infection as adverse effect of IL-1 blocking agent. Age-dependent difference in IL-1 signaling is not known.

The IL1B gene is conserved in chimpanzee, rhesus monkey, dog, cow, mouse, rat, and frog (<https://www.ncbi.nlm.nih.gov/homologene/481>), and the Myd88 gene is conserved in human, chimpanzee, rhesus monkey, dog, cow, rat, chicken, zebrafish, mosquito, and frog (https://www.ncbi.nlm.nih.gov/homologene?Db=homologene&Cmd=Retrieve&list_uids=1849).

These data suggest that the proposed AOP regarding inhibition of IL-1 signaling is not dependent on life stage, sex, age or species.

Key Event Description

1. Decreased IL-1 production

Decreased IL-1 production by macrophages or dendritic cells can be induced by suppressed IL-1 β mRNA induction or suppressed maturation of pro-IL-1 β . Dexamethasone is one of the representative drugs that significantly suppress IL-1 β production from monocytes (Finch-Arietta and Cochran, 1991). Other than dexamethasone, the inhibition of various targets in different layers from the stimulation of PRRs or the receptors of proinflammatory cytokines to the activation of NF- κ B or the inhibition of posttranscriptional regulation of pro-IL-1 β cause impaired IL-1R1 signaling. Among various PRRs, the signaling through TLR4 is best characterized. In addition, it is beyond the scope of this AOP to cover all signaling through each PRR. So, this AOP focuses on TLR4 signaling.

Binding of LPS to TLR4 and the coreceptor MD2 triggers interactions between the cytoplasmic TIR domain of TLR4 and TIR-containing adaptor proteins (Mal, MyD88, and TRAM). MyD88 binds IRAK4, which requires its kinase activity to bind the kinases IRAK1 and IRAK2 sequentially. The MyD88–IRAK complex also engages the ubiquitin ligase TRAF6 to make polyubiquitin chains that activate the IKK complex for NF- κ B- and ERK-dependent gene transcription. Ubiquitin ligases cIAP1 and cIAP2 recruited to the TLR4 signaling complex regulate translocation of a subset of signaling components to the cytoplasm, where TAK1 activation initiates a MAPK cascade, p38a and JNK, which stimulates gene expression. TLR4 activated at the plasma membrane is endocytosed but can signal within the endosomal compartment via the adaptors TRAM and TRIF. The kinase and ubiquitin ligase combination of RIP1 and Peli1 interacts with TRIF to signal NF- κ B activation, whereas TBK1 and TRAF3 stimulate IRF3-dependent transcription. Through these signaling cascades, NF- κ B, activator protein-1 (AP-1), cAMP responsive element binding protein (CREB)/ activating transcription factor (ATF), CCAAT-enhancer-binding protein b (c/EBP b), and interferon regulatory factor 3 (IRF3) are activated. These transcription factors induce the expression of various inflammatory cytokines e.g., IL-1 β , TNF α , IL-6 and several chemokines (reviewed by Newton and Dixit (Newton and Dixit, 2012)).

Therefore, chemicals that affect the signaling pathway leading to the activation of these transcription factors are supposed to suppress IL-1 β production. Among them, the chemical substances that affect NF- κ B signaling have been investigated most thoroughly. Quite a few compounds have been reported to inhibit NF- κ B signaling by several different mechanisms reviewed by Fuchs (Fuchs, 2010). In fact, dimethyl fumarate inhibits the activation of NF- κ B, resulting in a loss of proinflammatory cytokine production, distorted maturation and function of antigen-presenting cells, and immune deviation of T helper cells (Th) from the type 1 (Th1) and type 17 (Th17) profiles to a type 2 (Th2) phenotype (McGuire et al., 2016; Peng et al., 2012). Several studies have shown intriguing pharmacologic effects associated with curcumin, which inhibits NF- κ B expression by regulating NF- κ B/I κ B pathway and down-regulates expression of pro-inflammatory cytokines, such as IL-1, IL-6, IL-8, and TNF α (Wang et al., 2018). Iguratimod, a methanesulfonanilide, that is a novel disease-modifying antirheumatic drug, inhibits NF- κ B but not its inhibitor, I κ B α , and inhibits the production of IL-1 β (Mucke, 2012). Epigallocatechin gallate (EGCG) has been reported to inhibit NF- κ B activation through inhibition of p65 phosphorylation (Wheeler et al., 2004) and suppress the production of LPS-stimulated IL-1 β (Wang et al., 2020). DHMEQ inhibits LPS-induced NF- κ B activation by inhibiting its nuclear translocation from the cytoplasm. It also inhibits LPS-induced secretion of IL-1 β (Suzuki and Umezawa, 2006).

Other than the inhibitors for NF- κ B signaling, which can be stimulated by various stimulations other than TLR4 stimulation, there are signaling molecules that are specific to TLR4 signaling, such as TLR4, Mal, TRAM, Myd88, IRAK4, and IRAK1/2 (Vallabhapurapu and Karin, 2009). There are several chemicals that target some of these molecules, an inhibitors of TLR4 such as TAK-242 (Matsunaga et al., 2011) and various IRAK4 inhibitors (Lee et al., 2017). IRAK4 has recently attracted attention as a therapeutic target for inflammation and tumor diseases.

Beside transcriptional regulation of IL-1 β production, minocycline, and two prodrugs, pralnacasan (VX-740) and belnacasan (VX-765) that are orally absorbed and converted into the active principle, VRT-018858 and VRT-043198, respectively (Fenini et al., 2017) suppress IL-1 signaling by the inhibition of caspase-1 activation. Caspase-1 is an essential enzyme for maturation of pro-IL-1 β and the secretion of mature IL-1 β (Vincent and Mohr, 2007). Recently, it has been reported that cinnamaldehyde suppresses serum IL-1 β level in endotoxin poisoning mice (Xu et al., 2017).

2. Blocking of binding of IL-1 to IL-1R1

IL-1 α and IL-1 β independently bind the type I IL-1 receptor (IL-1R1), which is ubiquitously expressed. IL-1Ra binds IL-1R but does not initiate IL-1 signal transduction (Dripps et al., 1991). Recombinant IL-1Ra (anakinra) is fully active in blocking the IL-1R1, and therefore, the biological activities of IL-1 α and IL-1 β . The binding of IL-1 α and IL-1 β to IL-1R1 can be suppressed by soluble IL-1R like rilonacept (Kapur and Bonk, 2009). The binding of IL-1 β to IL-1R1 can be inhibited by anti-IL-1 β antibody (canakinumab and gevokizumab) (Church and McDermott, 2009) (Roell et al., 2010).

How it is Measured or Detected

1. Real time polymerase chain reaction to measure IL-1 α or IL-1 β mRNA
2. Enzyme-linked immunosorbent assay (ELISA) to detect IL-1 α or IL-1 β protein
3. Competitive inhibition binding experiments using 125 I-IL-1 α to type I IL-1R present on EL4 thymoma cells, 3T3 fibroblasts, hepatocytes, and Chinese hamster ovary cells expressing recombinant mouse type I IL-1R (McIntyre et al., 1991; Shuck et al., 1991).
4. Measure the ability of the reagent to neutralize the bioactivity of human IL-1 β on primary human fibroblasts in vitro (Alten et al., 2008)

References

- Alten, R., Gram, H., Joosten, L.A., et al., 2008. The human anti-IL-1 beta monoclonal antibody ACZ885 is effective in joint inflammation models in mice and in a proof-of-concept study in patients with rheumatoid arthritis. *Arthritis Res Ther* 10, R67.
- Chaudhary, D., Robinson, S., Romero, D.L., 2015. Recent advances in the discovery of small molecule inhibitors of interleukin-1 receptor-associated kinase 4 (IRAK4) as a therapeutic target for inflammation and oncology disorders. *J Med Chem* 58, 96-110.
- Church, L.D., McDermott, M.F., 2009. Canakinumab, a fully-human mAb against IL-1beta for the potential treatment of inflammatory disorders. *Curr Opin Mol Ther* 11, 81-89.
- Dripps, D.J., Brandhuber, B.J., Thompson, R.C., et al., 1991. Interleukin-1 (IL-1) receptor antagonist binds to the 80-kDa IL-1 receptor but does not initiate IL-1 signal transduction. *J Biol Chem* 266, 10331-10336.
- Fenini, G., Contassot, E., French, L.E., 2017. Potential of IL-1, IL-18 and Inflammasome Inhibition for the Treatment of Inflammatory Skin Diseases. *Front Pharmacol* 8, 278.
- Finch-Arietta, M.B., Cochran, F.R., 1991. Cytokine production in whole blood ex vivo. *Agents Actions* 34, 49-52.
- Fuchs, O., 2010. Transcription factor NF-kappaB inhibitors as single therapeutic agents or in combination with classical chemotherapeutic agents for the treatment of hematologic malignancies. *Curr Mol Pharmacol* 3, 98-122.
- Ho, S.C., Chang, Y.H., Chang, K.S., 2018. Structural Moieties Required for Cinnamaldehyde-Related Compounds to Inhibit Canonical IL-1beta Secretion. *Molecules* 23.
- Huang, H., Wang, Y., 2017. The protective effect of cinnamaldehyde on lipopolysaccharide induced acute lung injury in mice. *Cell Mol Biol (Noisy-le-grand)* 63, 58-63.
- Jeon, Y.J., Han, S.H., Lee, Y.W., et al., 2000. Dexamethasone inhibits IL-1 beta gene expression in LPS-stimulated RAW 264.7 cells by blocking NF-kappa B/Rel and AP-1 activation. *Immunopharmacology* 48, 173-183.
- Kapur, S., Bonk, M.E., 2009. Rilonacept (arcylast), an interleukin-1 trap for the treatment of cryopyrin-associated periodic syndromes. *P t* 34, 138-141.
- Klein, S.L., Flanagan, K.L., 2016. Sex differences in immune responses. *Nat Rev Immunol* 16, 626-638.
- Lee, K.L., Ambler, C.M., Anderson, D.R., et al., 2017. Discovery of Clinical Candidate 1-[[[(2S,3S,4S)-3-Ethyl-4-fluoro-5-oxopyrrolidin-2-yl]methoxy]-7-methoxyisoquinoline-6-carboxamide (PF-06650833), a Potent, Selective Inhibitor of Interleukin-1 Receptor Associated Kinase 4 (IRAK4), by Fragment-Based Drug Design. *J Med Chem* 60, 5521-5542.
- Matsunaga, N., Tsuchimori, N., Matsumoto, T., et al., 2011. TAK-242 (resatorvid), a small-molecule inhibitor of Toll-like receptor (TLR) 4 signaling, binds selectively to TLR4 and interferes with interactions between TLR4 and its adaptor molecules. *Mol Pharmacol* 79, 34-41.
- McGuire, V.A., Ruiz-Zorrilla Diez, T., Emmerich, C.H., et al., 2016. Dimethyl fumarate blocks pro-inflammatory cytokine production via inhibition of TLR induced M1 and K63 ubiquitin chain formation. *Sci Rep* 6, 31159.
- McIntyre, K.W., Stepan, G.J., Kolinsky, K.D., et al., 1991. Inhibition of interleukin 1 (IL-1) binding and bioactivity in vitro and modulation of acute inflammation in vivo by IL-1 receptor antagonist and anti-IL-1 receptor monoclonal antibody. *J Exp Med* 173, 931-939.
- Morand, E.F., Rickard, D., Goulding, N.J., 1993. Lack of involvement of lipocortin 1 in dexamethasone suppression of IL-1 release. *Mediators Inflamm* 2, 49-52.
- Mucke, H.A., 2012. Igaratimod: a new disease-modifying antirheumatic drug. *Drugs Today (Barc)* 48, 577-586.

- Newton, K., Dixit, V.M., 2012. Signaling in innate immunity and inflammation. *Cold Spring Harb Perspect Biol* 4.
- Peng, H., Guerau-de-Arellano, M., Mehta, V.B., et al., 2012. Dimethyl fumarate inhibits dendritic cell maturation via nuclear factor kappaB (NF-kappaB) and extracellular signal-regulated kinase 1 and 2 (ERK1/2) and mitogen stress-activated kinase 1 (MSK1) signaling. *J Biol Chem* 287, 28017-28026.
- Qiu, H.B., Pan, J.Q., Zhao, Y.Q., et al., 1997. Effects of dexamethasone and ibuprofen on LPS-induced gene expression of TNF alpha, IL-1 beta, and MIP-1 alpha in rat lung. *Zhongguo Yao Li Xue Bao* 18, 165-168.
- Quartier, P., 2011. Interleukin-1 antagonists in the treatment of autoinflammatory syndromes, including cryopyrin-associated periodic syndrome. *Open Access Rheumatol* 3, 9-18.
- Ravizza, T., Lucas, S.M., Balosso, S., et al., 2006. Inactivation of caspase-1 in rodent brain: a novel anticonvulsive strategy. *Epilepsia* 47, 1160-1168.
- Roell, M.K., Issafras, H., Bauer, R.J., et al., 2010. Kinetic approach to pathway attenuation using XOMA 052, a regulatory therapeutic antibody that modulates interleukin-1beta activity. *J Biol Chem* 285, 20607-20614.
- Shuck, M.E., Eessalu, T.E., Tracey, D.E., et al., 1991. Cloning, heterologous expression and characterization of murine interleukin 1 receptor antagonist protein. *Eur J Immunol* 21, 2775-2780.
- Stack, J.H., Beaumont, K., Larsen, P.D., et al., 2005. IL-converting enzyme/caspase-1 inhibitor VX-765 blocks the hypersensitive response to an inflammatory stimulus in monocytes from familial cold autoinflammatory syndrome patients. *J Immunol* 175, 2630-2634.
- Suzuki, E., Umezawa, K., 2006. Inhibition of macrophage activation and phagocytosis by a novel NF-kappaB inhibitor, dehydroxymethylepoxyquinomicin. *Biomed Pharmacother* 60, 578-586.
- Vallabhapurapu, S., Karin, M., 2009. Regulation and function of NF-kappaB transcription factors in the immune system. *Annu Rev Immunol* 27, 693-733.
- van Furth, A.M., Seijmonsbergen, E.M., Langermans, J.A., et al., 1995. Effect of xanthine derivatives and dexamethasone on *Streptococcus pneumoniae*-stimulated production of tumor necrosis factor alpha, interleukin-1 beta (IL-1 beta), and IL-10 by human leukocytes. *Clin Diagn Lab Immunol* 2, 689-692.
- Vincent, J.A., Mohr, S., 2007. Inhibition of caspase-1/interleukin-1beta signaling prevents degeneration of retinal capillaries in diabetes and galactosemia. *Diabetes* 56, 224-230.
- Wang, A.L., Yu, A.C., Lau, L.T., et al., 2005. Minocycline inhibits LPS-induced retinal microglia activation. *Neurochem Int* 47, 152-158.
- Wang, F., Han, Y., Xi, S., et al., 2020. Catechins reduce inflammation in lipopolysaccharide-stimulated dental pulp cells by inhibiting activation of the NF-kappaB pathway. *Oral Dis* 26, 815-821.
- Wang, Y., Tang, Q., Duan, P., et al., 2018. Curcumin as a therapeutic agent for blocking NF-kappaB activation in ulcerative colitis. *Immunopharmacol Immunotoxicol* 40, 476-482.
- Weinberg, J.B., Mason, S.N., Wortham, T.S., 1992. Inhibition of tumor necrosis factor-alpha (TNF-alpha) and interleukin-1 beta (IL-1 beta) messenger RNA (mRNA) expression in HL-60 leukemia cells by pentoxifylline and dexamethasone: dissociation of acivicin-induced TNF-alpha and IL-1 beta mRNA expression from acivicin-induced monocytoid differentiation. *Blood* 79, 3337-3343.
- Wheeler, D.S., Catravas, J.D., Odoms, K., et al., 2004. Epigallocatechin-3-gallate, a green tea-derived polyphenol, inhibits IL-1 beta-dependent proinflammatory signal transduction in cultured respiratory epithelial cells. *J Nutr* 134, 1039-1044.
- Xu, F., Wang, F., Wen, T., et al., 2017. Inhibition of NLRP3 inflammasome: a new protective mechanism of cinnamaldehyde in endotoxin poisoning of mice. *Immunopharmacol Immunotoxicol* 39, 296-304.

List of Key Events in the AOP

[Event: 202: Inhibition, Nuclear factor kappa B \(NF-kB\)](#)

Short Name: Inhibition, Nuclear factor kappa B (NF-kB)

Key Event Component

Process	Object	Action
I-kappaB kinase/NF-kappaB signaling	transcription factor NF-kappa-B subunit	decreased

AOPs Including This Key Event

AOP ID and Name	Event Type
Aop:14 - Glucocorticoid Receptor Activation Leading to Increased Disease Susceptibility	KeyEvent
Aop:278 - IKK complex inhibition leading to liver injury	KeyEvent
Aop:277 - Impaired IL-1R1 signaling leading to increased susceptibility to infection	KeyEvent

Stressors

Name
IL-1 receptor antagonist IL-1Ra (Anakinra)
anti-IL-1b antibody (Canakinumab)
soluble IL-1R (Rilonacept)

Biological Context

Level of Biological Organization

Molecular

Cell term

Cell term

macrophage

Organ term

Organ term

immune system

Domain of Applicability

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
Homo sapiens	Homo sapiens	High	NCBI
Mus musculus	Mus musculus	High	NCBI
Rattus norvegicus	Rattus norvegicus	High	NCBI

Life Stage Applicability

Life Stage Evidence

All life stages High

Sex Applicability

Sex Evidence

Unspecific High

The binding of sex steroids to their respective steroid receptors directly influences NF- κ B signaling, resulting in differential production of cytokines and chemokines (McKay and Cidlowski, 1999; Pernis, 2007). 17 β -estradiol regulates pro-inflammatory responses that are transcriptionally mediated by NF- κ B through a negative feedback and/or transrepressive interaction with NF- κ B (Straub, 2007). Progesterone suppresses innate immune responses and NF- κ B signal transduction reviewed by Klein et al. (Klein and Flanagan, 2016). Androgen-receptor signaling antagonises transcriptional factors NF- κ B (McKay and Cidlowski, 1999).

Evidence for perturbation of this molecular initiating event by stressor

Dex inhibits IL-1 β gene expression in LPS-stimulated RAW 264.7 cells by blocking NF- κ B/Rel and AP-1 activation (Jeon et al., 2000).

Various inhibitors for NF- κ B, such as dimethyl fumarate, curcumin, iguratimod, epigallocatechin gallate (EGCG), and DHMEQ inhibits LPS-induced NF- κ B activation and LPS-induced secretion of IL-1 β (McGuire et al., 2016; Mucke, 2012; Peng et al., 2012; Suzuki and Umezawa, 2006; Wang et al., 2020; Wang et al., 2018; Wheeler et al., 2004).

TAK-242 (Matsunaga et al., 2011) inhibit TLR4 itself. There are several IRAK4 inhibitors (Lee et al., 2017). These molecules block the upstream signal to NF- κ B activation. IRAK4 has recently attracted attention as a therapeutic target for inflammation and tumor diseases (Chaudhary et al., 2015).

LPS treatment induced a significant upregulation of the mRNA and release of IL-1 β from retinal microglia. Minocycline inhibited its releases. Thus, minocycline might exert its antiinflammatory effect on microglia by inhibiting the expression and release of IL-1 β (Wang et al., 2005).

Caspase-1 inhibition reduced the release of IL-1 β in organotypic slices exposed to LPS+ATP. Administration of pralnacasan (intracerebroventricular, 50 μ g) or belnacasan (intraperitoneal, 25–200 mg/kg) to rats blocked seizure-induced production of IL-1 β in the hippocampus, and resulted in a twofold delay in seizure onset and 50% reduction in seizure duration (Ravizza et al., 2006).

Belnacasan, an orally active IL-1 β converting enzyme/caspase-1 inhibitor, blocked IL-1 β secretion with equal potency in LPS-stimulated cells from familial cold urticarial associated syndrome and control subjects (Stack et al., 2005).

In LPS-induced acute lung injury (ALI) mice model, LPS induced inflammatory cytokines such as TNF- α , IL-6, IL-13 and IL-1 β were significantly decreased by cinnamaldehyde (CA) (Huang and Wang, 2017).

The suppressing capacities of six cinnamaldehyde-related compounds were evaluated and compared by using the LPS-primed and ATP-activated macrophages. At concentrations of 25~100 mM, cinnamaldehyde and 2-methoxy cinnamaldehyde dose-dependently inhibited IL-1 β secretion (Ho et al., 2018).

In vitro, CA decreased the levels of pro-IL-1 β and IL-1 β in cell culture supernatants, as well as the expression of NLRP3 and IL-1 β mRNA in cells. In vivo, CA decreased IL-1 β production in serum. Furthermore, CA suppressed LPS-induced NLRP3, p20, Pro-IL-1 β , P2X7 receptor (P2X7R) and cathepsin B protein expression in lung, as well as the expression of NLRP3 and IL-1 β mRNA (Xu et al., 2017).

IL-1Ra binds IL-1R but does not initiate IL-1 signal transduction (Dripps et al., 1991). Recombinant IL-1Ra (anakinra) is fully active in blocking the IL-1R1, and therefore, the biological activities of IL-1 α and IL-1 β . The binding of IL-1 α and IL-1 β to IL-1R1 can be suppressed by soluble IL-1R like rilonacept (Kapur and Bonk, 2009). The binding of IL-1 β to IL-1R1 can be inhibited by anti-IL-1 β antibody (canakinumab and gevokizumab) (Church and McDermott, 2009) (Roell et al., 2010).

IL-1 is known to mediate autoinflammatory syndrome, such as cryopyrin-associated periodic syndrome, neonatal-onset multisystem inflammatory disease and familial Mediterranean fever. Blocking of binding of IL-1 to IL-1R1 by anakinra, canakinumab, and rilonacept have been already used to treat these autoinflammatory syndrome associated with overactivation of IL-1 signaling (Quartier, 2011).

Dex inhibits IL-1 β gene expression in LPS-stimulated RAW 264.7 cells by blocking NF- κ B/Rel and AP-1 activation (Jeon et al., 2000).

Inhibition of IL-1 binding to IL-1R or the decreased production of IL-1b leads to the suppression of IL-1R signaling leading to NF- κ B activation.

Key Event Description

The NF- κ B pathway consists of a series of events where the transcription factors of the NF- κ B family play the key role. The canonical NF- κ B pathway can be activated by a range of stimuli, including TNF receptor activation by TNF- α . Upon pathway activation, the IKK complex will be phosphorylated, which in turn phosphorylates I κ B α . This NF- κ B inhibitor will be K48-linked ubiquitinated and degraded, allowing NF- κ B to translocate to the nucleus. There, this transcription factor can express pro-inflammatory and anti-apoptotic genes. Furthermore, negative feedback genes are also transcribed and include I κ B α and A20. When the NF- κ B pathway is inhibited, its translocation will be delayed (or absent), resulting in less or no regulation of NF- κ B target genes. This can be achieved by IKK inhibitors, proteasome inhibitors, nuclear translocation inhibitors or DNA-binding inhibitors (Gupta et al., 2010; Liu et al., 2017). Therefore, inhibition of IL-1R activation suppresses NF- κ B.

How it is Measured or Detected

NF- κ B transcriptional activity: Beta lactamase reporter gene assay (Miller et al. 2010)

NF- κ B transcription: Lentiviral NF- κ BGFP reporter with flow cytometry (Moujalled et al. 2012)

I κ B α phosphorylation: Western blotting (Miller et al. 2010)

NF- κ B p65 (Total/Phospho) ELISA

ELISA for IL-6, IL-8, and Cox

References

- Chaudhary, D., Robinson, S., Romero, D.L. (2015), Recent advances in the discovery of small molecule inhibitors of interleukin-1 receptor-associated kinase 4 (IRAK4) as a therapeutic target for inflammation and oncology disorders. *J Med Chem* 58: 96-110, 10.1021/jm5016044
- Church, L.D., McDermott, M.F. (2009), Canakinumab, a fully-human mAb against IL-1beta for the potential treatment of inflammatory disorders. *Curr Opin Mol Ther* 11: 81-89,
- Dripps, D.J., Brandhuber, B.J., Thompson, R.C., et al. (1991), Interleukin-1 (IL-1) receptor antagonist binds to the 80-kDa IL-1 receptor but does not initiate IL-1 signal transduction. *J Biol Chem* 266: 10331-10336,
- Gupta, S.C., Sundaram, C., Reuter, S., et al. (2010), Inhibiting NF-kappaB activation by small molecules as a therapeutic strategy. *Biochim Biophys Acta* 1799: 775-787, 10.1016/j.bbagr.2010.05.004
- Ho, S.C., Chang, Y.H., Chang, K.S. (2018), Structural Moieties Required for Cinnamaldehyde-Related Compounds to Inhibit Canonical IL-1beta Secretion. *Molecules* 23, 10.3390/molecules23123241
- Huang, H., Wang, Y. (2017), The protective effect of cinnamaldehyde on lipopolysaccharide induced acute lung injury in mice. *Cell Mol Biol (Noisy-le-grand)* 63: 58-63, 10.14715/cmb/2017.63.8.13
- Jeon, Y.J., Han, S.H., Lee, Y.W., et al. (2000), Dexamethasone inhibits IL-1 beta gene expression in LPS-stimulated RAW 264.7 cells by blocking NF-kappa B/Rel and AP-1 activation. *Immunopharmacology* 48: 173-183,
- Kapur, S., Bonk, M.E. (2009), Riloncept (arcalyst), an interleukin-1 trap for the treatment of cryopyrin-associated periodic syndromes. *P t* 34: 138-141,
- Klein, S.L., Flanagan, K.L. (2016), Sex differences in immune responses. *Nat Rev Immunol* 16: 626-638, 10.1038/nri.2016.90
- Lee, K.L., Ambler, C.M., Anderson, D.R., et al. (2017), Discovery of Clinical Candidate 1-[(2S,3S,4S)-3-Ethyl-4-fluoro-5-oxopyrrolidin-2-yl]methoxy}-7-methoxyisoquinoline-6-carboxamide (PF-06650833), a Potent, Selective Inhibitor of Interleukin-1 Receptor Associated Kinase 4 (IRAK4), by Fragment-Based Drug Design. *J Med Chem* 60: 5521-5542, 10.1021/acs.jmedchem.7b00231
- Liu, T., Zhang, L., Joo, D., et al. (2017), NF-kappaB signaling in inflammation. *Signal Transduct Target Ther* 2, 10.1038/sigtrans.2017.23
- Matsunaga, N., Tsuchimori, N., Matsumoto, T., et al. (2011), TAK-242 (resatorvid), a small-molecule inhibitor of Toll-like receptor (TLR) 4 signaling, binds selectively to TLR4 and interferes with interactions between TLR4 and its adaptor molecules. *Mol Pharmacol* 79: 34-41, 10.1124/mol.110.068064
- McGuire, V.A., Ruiz-Zorrilla Diez, T., Emmerich, C.H., et al. (2016), Dimethyl fumarate blocks pro-inflammatory cytokine production via inhibition of TLR induced M1 and K63 ubiquitin chain formation. *Sci Rep* 6: 31159, 10.1038/srep31159
- McKay, L.I., Cidlowski, J.A. (1999), Molecular control of immune/inflammatory responses: interactions between nuclear factor-kappa B and steroid receptor-signaling pathways. *Endocr Rev* 20: 435-459, 10.1210/edrv.20.4.0375
- Mucke, H.A. (2012), Iguratimod: a new disease-modifying antirheumatic drug. *Drugs Today (Barc)* 48: 577-586, 10.1358/dot.2012.48.9.1855758
- Peng, H., Guerau-de-Arellano, M., Mehta, V.B., et al. (2012), Dimethyl fumarate inhibits dendritic cell maturation via nuclear factor kappaB (NF-kappaB) and extracellular signal-regulated kinase 1 and 2 (ERK1/2) and mitogen stress-activated kinase 1 (MSK1) signaling. *J Biol Chem* 287: 28017-28026, 10.1074/jbc.M112.383380
- Pernis, A.B. (2007), Estrogen and CD4+ T cells. *Curr Opin Rheumatol* 19: 414-420, 10.1097/BOR.0b013e328277ef2a
- Quartier, P. (2011), Interleukin-1 antagonists in the treatment of autoinflammatory syndromes, including cryopyrin-associated periodic syndrome. *Open Access Rheumatol* 3: 9-18, 10.2147/oarr.S6696
- Ravizza, T., Lucas, S.M., Balosso, S., et al. (2006), Inactivation of caspase-1 in rodent brain: a novel anticonvulsive strategy. *Epilepsia* 47: 1160-1168, 10.1111/j.1528-1167.2006.00590.x
- Roell, M.K., Issafras, H., Bauer, R.J., et al. (2010), Kinetic approach to pathway attenuation using XOMA 052, a regulatory therapeutic antibody that modulates interleukin-1beta activity. *J Biol Chem* 285: 20607-20614, 10.1074/jbc.M110.115790
- Stack, J.H., Beaumont, K., Larsen, P.D., et al. (2005), IL-converting enzyme/caspase-1 inhibitor VX-765 blocks the hypersensitive response to an inflammatory stimulus in monocytes from familial cold autoinflammatory syndrome patients. *J Immunol* 175: 2630-2634,
- Straub, R.H. (2007), The complex role of estrogens in inflammation. *Endocr Rev* 28: 521-574, 10.1210/er.2007-0001

Suzuki, E., Umezawa, K. (2006), Inhibition of macrophage activation and phagocytosis by a novel NF-kappaB inhibitor, dehydroxymethylepoxyquinomicin. *Biomed Pharmacother* 60: 578-586, 10.1016/j.biopha.2006.07.089

Wang, A.L., Yu, A.C., Lau, L.T., et al. (2005), Minocycline inhibits LPS-induced retinal microglia activation. *Neurochem Int* 47: 152-158, 10.1016/j.neuint.2005.04.018

Wang, F., Han, Y., Xi, S., et al. (2020), Catechins reduce inflammation in lipopolysaccharide-stimulated dental pulp cells by inhibiting activation of the NF-kappaB pathway. *Oral Dis* 26: 815-821, 10.1111/odi.13290

Wang, Y., Tang, Q., Duan, P., et al. (2018), Curcumin as a therapeutic agent for blocking NF-kappaB activation in ulcerative colitis. *Immunopharmacol Immunotoxicol* 40: 476-482, 10.1080/08923973.2018.1469145

Wheeler, D.S., Catravas, J.D., Odoms, K., et al. (2004), Epigallocatechin-3-gallate, a green tea-derived polyphenol, inhibits IL-1 beta-dependent proinflammatory signal transduction in cultured respiratory epithelial cells. *J Nutr* 134: 1039-1044, 10.1093/jn/134.5.1039

Xu, F., Wang, F., Wen, T., et al. (2017), Inhibition of NLRP3 inflammasome: a new protective mechanism of cinnamaldehyde in endotoxin poisoning of mice. *Immunopharmacol Immunotoxicol* 39: 296-304, 10.1080/08923973.2017.1355377

[Event: 1702: Suppression of T cell activation](#)

Short Name: Suppression of T cell activation

AOPs Including This Key Event

AOP ID and Name	Event Type
Aop:277 - Impaired IL-1R1 signaling leading to increased susceptibility to infection	KeyEvent

Biological Context

Level of Biological Organization

Cellular

Cell term

Cell term

T cell

Organ term

Organ term

immune system

Domain of Applicability

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
Homo sapiens	Homo sapiens	High	NCBI
Mus musculus	Mus musculus	High	NCBI
Rattus norvegicus	Rattus norvegicus	High	NCBI

Life Stage Applicability

Life Stage Evidence

All life stages High

Sex Applicability

Sex Evidence

Unspecific High

Key Event Description

T cells are key orchestrators of the response against pathogens and are also fundamental in maintaining self-tolerance. A number of clinically important conditions have been described in which T-cell functions are altered, as in AIDS or upon immunosuppression after application of various immunosuppressive drugs to treat autoimmune disorders or allogeneic graft rejection. T-cell progenitors differentiate in the thymus into immature T cells that acquire the expression of the T-cell receptor (TCR), which recognizes antigen peptides from pathogens presented along with major histocompatibility complex (MHC). In addition to the TCR, T cells are characterized by expression of the co-receptor molecules CD4 and CD8 on their cell surface. CD4+ T cells, also called T helper (Th) cells, recognize antigen/MHC-II complexes on antigen presenting cells (APCs) and coordinate the activation of other immune cells including B cells, macrophages, etc.

Therefore, CD4+ T cells are crucial for coordination of the immune response and for the elimination of invading pathogens. On the other hand, CD8+ T cells, referred to as T cytotoxic cells, recognize antigen/MHC-I complexes and are responsible for the killing of pathogen-infected cells.

T-cell activation and differentiation depends on antigen presenting cells (APCs) such as dendritic cells (DCs), macrophages and B cells. depending on the insult affecting a given tissue. Different subsets of DCs can be generated that in turn are able to coordinate the differentiation of a particular Th subset. To date, the following Th subsets have been described: Th1, Th2, Th9, Th17, Th22, Tfh (follicular helper T cells), Tr1 (type 1 regulatory T cells) and Treg (regulatory T cells), each possessing a specific function in the elimination of pathogens. (reviewed by Simeoni et al. (Simeoni et al., 2016))

Although CD4 T cells are able to commit to Th1, Th2 and Th17 lineages in the absence of IL-1R signaling at steady state, these committed CD4 T cells are unable to effectively secrete their cytokines upon TCR ligation. Namely, IL-1 is indispensable for CD4 T cell effector function. (Lin et al, 2015)

Moreover, since full activation of B cells and antibody production and class switch depends on T cell help. The impaired activation of T cells leads to impaired B cell activation and antibody production (reviewed by Mok (Mok, 2010)).

How it is Measured or Detected

T cell activation can be evaluated by measuring IL-2 production by ELISA or T cell proliferation by incorporation of the analysis of CFSE labeled T cells or [³H]thymidine incorporation.

References

- Lin, D., Lei, L., Zhang, Y., et al. (2015), Secreted IL-1α promotes T-cell activation and expansion of CD11b(+) Gr1(+) cells in carbon tetrachloride-induced liver injury in mice. *Eur J Immunol* 45: 2084-2098, 10.1002/eji.201445195
- Mok, M.Y. (2010), The immunological basis of B-cell therapy in systemic lupus erythematosus. *Int J Rheum Dis* 13: 3-11, 10.1111/j.1756-185X.2009.01458.x
- Simeoni, L., Thurm, C., Kritikos, A., et al. (2016), Redox homeostasis, T cells and kidney diseases: three faces in the dark. *Clin Kidney J* 9: 1-10, 10.1093/ckj/sfv135
- Weih, F., Carrasco, D., Durham, S.K., et al. (1995), Multiorgan inflammation and hematopoietic abnormalities in mice with a targeted disruption of RelB, a member of the NF-κB/Rel family. *Cell* 80: 331-340,

List of Adverse Outcomes in this AOP

[Event: 986: Increase, Increased susceptibility to infection](#)

Short Name: Increase, Increased susceptibility to infection

AOPs Including This Key Event**AOP ID and Name****Event Type**

[Aop:277 - Impaired IL-1R1 signaling leading to increased susceptibility to infection](#) AdverseOutcome

Stressors

Name

IL-1 receptor antagonist IL-1Ra (Anakinra)
 anti-IL-1b antibody (Canakinumab)
 soluble IL-1R (Rilonacept)

Biological Context**Level of Biological Organization**

Individual

Domain of Applicability**Taxonomic Applicability**

Term	Scientific Term	Evidence	Links
Homo sapiens	Homo sapiens	High	NCBI
Mus musculus	Mus musculus	High	NCBI
Rattus norvegicus	Rattus norvegicus	High	NCBI

Life Stage Applicability**Life Stage Evidence**

All life stages High

Sex Applicability**Sex Evidence**

Unspecific High

The increased susceptibility to infection caused by IL-1RA or anti-IL-1 antibody has been reported in both humans and mice (De Benedetti et al., 2018; Fleischmann et al., 2003).

Key Event Description

Severe combined immunodeficiencies (SCIDs) comprise a group of rare, monogenic diseases that are characterized by an early onset and a profound block in the development of T lymphocytes. Given that adaptive immunity is abrogated, patients with SCID are prone to recurrent infections caused by both non-opportunistic and opportunistic pathogens, leading to early death unless immunity can be restored reviewed by Fischer et al. (Fischer et al., 2015). Human immunodeficiency virus (HIV) is a retrovirus known to attack the CD4+ T lymphocytes. In individuals with chronic HIV infection not on treatment with antiretroviral agents, as the CD4+ count drops they are vulnerable to a multitude of infections which rarely occur in an immunocompetent host, hence the term opportunistic infections (Justiz Vaillant and Naik, 2021). Various immunosuppressive agents such as corticosteroids, antimetabolites, calcineurin inhibitors, glucocorticoids, antithymocyte globulin, antibodies against IL-2RA or CD28, which all suppress T cell function, increase the incidence of opportunistic infection (reviewed by Tasdogan et al. (Tasdogan et al., 2019).

How it is Measured or Detected

By comparison of the incidence of infection between individuals exposed to stressors and non-exposed individuals.

Regulatory Significance of the AO

Increased susceptibility to infection is the significant adverse outcome of drugs and chemicals present in the environment. Therefore, drugs or chemicals that have an effect that can cause immunosuppression must be under regulatory scrutiny.

References

De Benedetti, F., Gattorno, M., Anton, J., et al. (2018), Canakinumab for the Treatment of Autoinflammatory Recurrent Fever Syndromes. *N Engl J Med* 378: 1908-1919, 10.1056/NEJMoa1706314

Fang, X.M., Schroder, S., Hoefft, A., et al. (1999), Comparison of two polymorphisms of the interleukin-1 gene family: interleukin-1 receptor antagonist polymorphism contributes to susceptibility to severe sepsis. *Crit Care Med* 27: 1330-1334, 10.1097/00003246-199907000-00024

- Fischer, A., Notarangelo, L.D., Neven, B., et al. (2015), Severe combined immunodeficiencies and related disorders. *Nat Rev Dis Primers* 1: 15061, 10.1038/nrdp.2015.61
- Fleischmann, R.M., Schechtman, J., Bennett, R., et al. (2003), Anakinra, a recombinant human interleukin-1 receptor antagonist (r-methHuL-1ra), in patients with rheumatoid arthritis: A large, international, multicenter, placebo-controlled trial. *Arthritis Rheum* 48: 927-934, 10.1002/art.10870
- Genovese, M.C., Cohen, S., Moreland, L., et al. (2004), Combination therapy with etanercept and anakinra in the treatment of patients with rheumatoid arthritis who have been treated unsuccessfully with methotrexate. *Arthritis Rheum* 50: 1412-1419, 10.1002/art.20221
- Guler, R., Parihar, S.P., Spohn, G., et al. (2011), Blocking IL-1alpha but not IL-1beta increases susceptibility to chronic Mycobacterium tuberculosis infection in mice. *Vaccine* 29: 1339-1346, 10.1016/j.vaccine.2010.10.045
- Horino, T., Matsumoto, T., Ishikawa, H., et al. (2009), Interleukin-1 deficiency in combination with macrophage depletion increases susceptibility to Pseudomonas aeruginosa bacteremia. *Microbiol Immunol* 53: 502-511, 10.1111/j.1348-0421.2009.00143.x
- Imagawa, T., Nishikomori, R., Takada, H., et al. (2013), Safety and efficacy of canakinumab in Japanese patients with phenotypes of cryopyrin-associated periodic syndrome as established in the first open-label, phase-3 pivotal study (24-week results). *Clin Exp Rheumatol* 31: 302-309,
- Juffermans, N.P., Florquin, S., Camoglio, L., et al. (2000), Interleukin-1 signaling is essential for host defense during murine pulmonary tuberculosis. *J Infect Dis* 182: 902-908, 10.1086/315771
- Justiz Vaillant, A.A., Naik, R. 2021. HIV-1 Associated Opportunistic Infections, *StatPearls*. StatPearls Publishing
- Copyright © 2021, StatPearls Publishing LLC., Treasure Island (FL).
- Kullenberg, T., Lofqvist, M., Leinonen, M., et al. (2016), Long-term safety profile of anakinra in patients with severe cryopyrin-associated periodic syndromes. *Rheumatology (Oxford)* 55: 1499-1506, 10.1093/rheumatology/kew208
- Lachmann, H.J., Kone-Paut, I., Kuemmerle-Deschner, J.B., et al. (2009), Use of canakinumab in the cryopyrin-associated periodic syndrome. *N Engl J Med* 360: 2416-2425, 10.1056/NEJMoa0810787
- Lequerre, T., Quartier, P., Rosellini, D., et al. (2008), Interleukin-1 receptor antagonist (anakinra) treatment in patients with systemic-onset juvenile idiopathic arthritis or adult onset Still disease: preliminary experience in France. *Ann Rheum Dis* 67: 302-308, 10.1136/ard.2007.076034
- Migkos, M.P., Somarakis, G.A., Markatseli, T.E., et al. (2015), Tuberculous pyomyositis in a rheumatoid arthritis patient treated with anakinra. *Clin Exp Rheumatol* 33: 734-736,
- Motsinger-Reif, A.A., Antas, P.R., Oki, N.O., et al. (2010), Polymorphisms in IL-1beta, vitamin D receptor Fok1, and Toll-like receptor 2 are associated with extrapulmonary tuberculosis. *BMC Med Genet* 11: 37, 10.1186/1471-2350-11-37
- Schlesinger, N., Alten, R.E., Bardin, T., et al. (2012), Canakinumab for acute gouty arthritis in patients with limited treatment options: results from two randomised, multicentre, active-controlled, double-blind trials and their initial extensions. *Ann Rheum Dis* 71: 1839-1848, 10.1136/annrheumdis-2011-200908
- Tasdogan, B.E., Ma, M., Simsek, C., et al. (2019), Update on Immunosuppression in Liver Transplantation. *Euroasian J Hepatogastroenterol* 9: 96-101, 10.5005/jp-journals-10018-1301
- Tian, T., Jin, M.Q., Dubin, K. (2017), IL-1R Type 1-Deficient Mice Demonstrate an Impaired Host Immune Response against Cutaneous Vaccinia Virus Infection. 198: 4341-4351, 10.4049/jimmunol.1500106
- Wojtowicz, A., Gresnigt, M.S., Lecompte, T., et al. (2015), IL1B and DEFB1 Polymorphisms Increase Susceptibility to Invasive Mold Infection After Solid-Organ Transplantation. *J Infect Dis* 211: 1646-1657, 10.1093/infdis/jiu636
- Yamada, H., Mizumo, S., Horai, R., et al. (2000), Protective role of interleukin-1 in mycobacterial infection in IL-1 alpha/beta double-knockout mice. *Lab Invest* 80: 759-767,
- Yokota, S., Imagawa, T., Nishikomori, R., et al. (2017), Long-term safety and efficacy of canakinumab in cryopyrin-associated periodic syndrome: results from an open-label, phase III pivotal study in Japanese patients. *Clin Exp Rheumatol* 35 Suppl 108: 19-26,

Appendix 2

List of Key Event Relationships in the AOP

List of Adjacent Key Event Relationships

Relationship: 2002: Impaired IL-1R1 signaling leads to Inhibition, Nuclear factor kappa B (NF-kB)

AOPs Referencing Relationship

AOP Name	Adjacency	Weight of Evidence	Quantitative Understanding
Impaired IL-1R1 signaling leading to increased susceptibility to infection	adjacent	High	Moderate

Evidence Supporting Applicability of this Relationship

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
Homo sapiens	Homo sapiens	High	NCBI
Mus musculus	Mus musculus	High	NCBI
Rattus norvegicus	Rattus norvegicus	High	NCBI

Life Stage Applicability

Life Stage	Evidence
All life stages	High

Sex Applicability

Sex	Evidence
Unspecific	High

Key Event Relationship Description

After binding of IL-1a or IL-1b to IL-1R, IL-1 and IL-1R1 facilitates recruitment of IL-1RacP. Then this trimeric complex rapidly assembles two intracellular signaling proteins, myeloid differentiation primary response gene 88 (MYD88) and interleukin-1 receptor-activated protein kinase (IRAK) 4. IL-1, IL-1RI, IL-RacP, MYD88, and IRAK4 form a stable IL-1-induced first signaling module. The binding of MyD88 triggers a cascade of kinases that produce a strong pro-inflammatory signal leading to activation of NF-κB.

Evidence Supporting this KER

Mice lacking MYD88 or IRAK4 show severe defects in IL-1 signaling (Adachi et al., 1998; Medzhitov et al., 1998; Suzuki et al., 2002). Similarly, humans with mutations in the IRAK4 gene have defects in IL-1RI and Toll-like receptor (TLR) signaling (Picard et al., 2003).

Biological Plausibility

The initial step in IL-1 signal transduction is a ligand-induced conformational change in the first extracellular domain of the IL-1RI that facilitates recruitment of IL-1RacP (Cavalli et al., 2015). Through conserved cytosolic regions called Toll- and IL-1R-like (TIR) domains (Radons et al., 2003), the trimeric complex rapidly assembles two intracellular signaling proteins, myeloid differentiation primary response gene 88 (MYD88) and interleukin-1 receptor-activated protein kinase (IRAK) 4 (Brikos et al., 2007; Li et al., 2002). IL-1, IL-1RI, IL-RacP, MYD88, and IRAK4 form a stable IL-1-induced first signaling module. The binding of MyD88 triggers a cascade of kinases that produce a strong pro-inflammatory signal leading to activation of NF-κB reviewed by (Brikos et al., 2007; Weber, Wasiliew and Kracht, 2010).

Empirical Evidence

IL-1Ra blocks IL-1 signaling:

IL-1Ra down modulation of EGF receptor (3 nM of ED50) (Dripps et al., 1991)

IL-1Ra suppression of IL-1-induced endothelial cell-leukocyte adhesion (approximately 10 ng/ml of ED50) (Dripps et al., 1991)

IL-1Ra suppresses rhIL-1a-induced mouse thymocytes proliferation (ED50 almost 3 mg/mL) (Arend et al., 1990)

IL-1Ra competed for binding of ¹²⁵I-IL-1a to type I IL-1R present on EL4 thymoma cells, 3T3 fibroblasts, hepatocytes, and Chinese hamster ovary cells expressing recombinant mouse type I IL-1R. The IC50 values for IL-1ra binding (ranging from 2 to 4

ng/ml) were similar to those of IL-1a. (McIntyre et al., 1991)

Recombinant mL-1Ra competitively inhibited ^{125}I -labeled IL-1 alpha binding to murine type I IL-1R present on EL4 6.1 cells (Ki value of 0.21 nM) and antagonized IL-1-stimulated co-mitogenesis in murine thymocytes ($0.7 \times 10(6)$ - $1.1 \times 10(6)$ units/mg). (Shuck et al., 1991)

Peripheral blood mononuclear cells (PBMC) obtained after completion of the IL-lra infusion synthesized significantly less interleukin 6 ex vivo than PBMC from saline-injected controls. (Granowitz et al., 1992)

Canakinumab (ACZ885, Ilaris):

Canakinumab binds to human IL-1 β with high affinity; the antibody-antigen dissociation equilibrium constant is approximately 35–40 pM (Dhimolea, 2010).

The antibody binds to human IL-1 β with high affinity (about 40 pmol/l). The antibody was found to neutralize the bioactivity of human IL-1 β on primary human fibroblasts in vitro 44.6 pmol/l (7.1 ± 0.56 ng/ml; n = 6) of ED50. Application of Canakinumab intraperitoneally 2 hours before injecting the IL-1 β producing cells completely suppressed joint swelling (0.06 mg/kg of EC50) (Alten et al., 2008).

Primary human fibroblasts are stimulated with recombinant IL-1b or conditioned medium obtained from LPS-stimulated human PBMCs in the presence of various concentrations of Canakinumab or IL-1RA ranging from 6 to 18,000 pM. Supernatant is taken after 16 h stimulation and assayed for IL-6 by ELISA. Canakinumab typically have 1 nM or less of EC50 for inhibition of IL-6 production (Canakinumab Patent Application WO02/16436.)

Rilonacept (IL-1 Trap, Arcalyst):

Incubation of the human MRC5 fibroblastic cell line with IL-1 β induces secretion of IL-6. At a constant amount of IL-1 β (4 pM), the IC50 of the IL-1 trap is ~2 pM. Another unique property of the IL-1 trap is that it not only blocks IL-1 β , but also blocks IL-1 α with high affinity (KD = ~3 pM; data not shown). The titration curve of IL-1 trap in the presence of 10 pM IL-1 β shows an IC50 of 6.5 pM, which corresponds to a calculated KD of 1.5 pM (This affinity is 100 times higher than that of the soluble single component receptor IL-1RI (Economides et al., 2003).

IRAK4 inhibitor

By reconstituting IRAK-4-deficient cells with wild type or kinase-inactive IRAK-4, it is demonstrated that the kinase activity of IRAK-4 is required for the optimal transduction of IL-1-induced signals, including the activation of IRAK-1, NF- κ B, and JNK, and the maximal induction of inflammatory cytokines (Lye et al., 2008).

Various concentrations of kinase-active or kinase-inactive IRAK-4 were transiently (Lye et al.) overexpressed in IRAK-4-deficient cells that were also transiently transfected with an NF- κ B-dependent luciferase reporter and α -galactosidase expression vector. Transfected cells were left untreated or treated with IL-1 β (10 ng/ml) for 6 h before luciferase and α -galactosidase activities were measured. The luciferase activity was divided by the α -galactosidase activity, and fold activation was calculated compared with the activity of untreated cells carrying an empty α -vector (normalized as 1). The results demonstrated that kinase-active IRAK-4 dose dependently activates NF- κ B (Lye et al., 2004).

Quantitative Understanding of the Linkage

See Empirical Evidence.

Response-response relationship

IL-1Ra blocks IL-1 signaling:

Suppression of IL-1-induced IL-1, TNFa, or IL-6 synthesis was dose-dependent ($P \leq .0001$). At a twofold molar excess, IL-lra inhibited IL-1-induced IL-1 or TNFa synthesis by 50% ($P < .01$); an equimolar concentration of IL-lra inhibited synthesis of these two cytokines by over 20% ($P < .05$). A 10-fold molar excess of IL-lra over IL-lb reduced IL-lb-induced IL-lra by 95% ($P = .01$) and IL-lra-induced IL-lb by 73% ($P < .01$). In elutriated monocytes, a 10-fold molar excess of IL-lra reduced IL-lb-induced IL-lra by 82% ($P < .05$), TNFa by 64% ($P = .05$), and IL-6 by 47% ($P < .05$). (Granowitz et al., 1992)

Rilonacept (IL-1 Trap, Arcalyst) blocks IL-1 signaling:

The titration curve of IL-1 trap in the presence of 10 pM IL-1 β shows an IC50 of 6.5 pM, which corresponds to a calculated KD of 1.5 pM (This affinity is 100 times higher than that of the soluble single component receptor IL-1RI (Economides et al., 2003).

References

Adachi, O., Kawai, T., Takeda, K., et al. (1998), Targeted disruption of the MyD88 gene results in loss of IL-1- and IL-18-mediated function. *Immunity* 9: 143-150,

- Alten, R., Gram, H., Joosten, L.A., et al. (2008), The human anti-IL-1 beta monoclonal antibody ACZ885 is effective in joint inflammation models in mice and in a proof-of-concept study in patients with rheumatoid arthritis. *Arthritis Res Ther* 10: R67, 10.1186/ar2438
- Arend, W.P., Welgus, H.G., Thompson, R.C., et al. (1990), Biological properties of recombinant human monocyte-derived interleukin 1 receptor antagonist. *J Clin Invest* 85: 1694-1697, 10.1172/jci114622
- Dhimolea, E. (2010), Canakinumab. *MAbs* 2: 3-13,
- Dripps, D.J., Brandhuber, B.J., Thompson, R.C., et al. (1991), Interleukin-1 (IL-1) receptor antagonist binds to the 80-kDa IL-1 receptor but does not initiate IL-1 signal transduction. *J Biol Chem* 266: 10331-10336,
- Economides, A.N., Carpenter, L.R., Rudge, J.S., et al. (2003), Cytokine traps: multi-component, high-affinity blockers of cytokine action. *Nat Med* 9: 47-52, 10.1038/nm811
- Granowitz, E.V., Clark, B.D., Vannier, E., et al. (1992), Effect of interleukin-1 (IL-1) blockade on cytokine synthesis: I. IL-1 receptor antagonist inhibits IL-1-induced cytokine synthesis and blocks the binding of IL-1 to its type II receptor on human monocytes. *Blood* 79: 2356-2363,
- Lye, E., Dhanji, S., Calzascia, T., et al. (2008), IRAK-4 kinase activity is required for IRAK-4-dependent innate and adaptive immune responses. *Eur J Immunol* 38: 870-876, 10.1002/eji.200737429
- Lye, E., Mirtsos, C., Suzuki, N., et al. (2004), The role of interleukin 1 receptor-associated kinase-4 (IRAK-4) kinase activity in IRAK-4-mediated signaling. *J Biol Chem* 279: 40653-40658, 10.1074/jbc.M402666200
- McIntyre, K.W., Stepan, G.J., Kolinsky, K.D., et al. (1991), Inhibition of interleukin 1 (IL-1) binding and bioactivity in vitro and modulation of acute inflammation in vivo by IL-1 receptor antagonist and anti-IL-1 receptor monoclonal antibody. *J Exp Med* 173: 931-939,
- Medzhitov, R., Preston-Hurlburt, P., Kopp, E., et al. (1998), MyD88 is an adaptor protein in the hToll/IL-1 receptor family signaling pathways. *Mol Cell* 2: 253-258,
- Picard, C., Puel, A., Bonnet, M., et al. (2003), Pyogenic bacterial infections in humans with IRAK-4 deficiency. *Science* 299: 2076-2079, 10.1126/science.1081902
- Shuck, M.E., Eessalu, T.E., Tracey, D.E., et al. (1991), Cloning, heterologous expression and characterization of murine interleukin 1 receptor antagonist protein. *Eur J Immunol* 21: 2775-2780, 10.1002/eji.1830211119
- Suzuki, N., Suzuki, S., Duncan, G.S., et al. (2002), Severe impairment of interleukin-1 and Toll-like receptor signalling in mice lacking IRAK-4. *Nature* 416: 750-756, 10.1038/nature736

Relationship: 2003: Inhibition, Nuclear factor kappa B (NF-kB) leads to Suppression of T cell activation

AOPs Referencing Relationship

AOP Name	Adjacency	Weight of Evidence	Quantitative Understanding
Impaired IL-1R1 signaling leading to increased susceptibility to infection	adjacent	High	Moderate

Evidence Supporting Applicability of this Relationship

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
Homo sapiens	Homo sapiens	High	NCBI
Mus musculus	Mus musculus	High	NCBI
Rattus norvegicus	Rattus norvegicus	High	NCBI

Life Stage Applicability

Life Stage	Evidence
All life stages	High

Sex Applicability

Sex	Evidence
Unspecific	High

Key Event Relationship Description

NF- κ B plays a crucial role in the activation of dendritic cells as well as T cells. In dendritic cells, the activation of the canonical NF- κ B pathway in response to pro-inflammatory stimuli, such as cytokines including IL-1 α or IL-1 β and TLR ligands, stimulate the maturation of dendritic cells with enhanced antigen presenting function. The inhibition of NF- κ B suppress antigen presenting function of dendritic cells, resulting in suppression of T cell activation (reviewed by Reinhard et al (Reinhard et al., 2012) and van Delft et al (van Delft, Huitema and Tas, 2015)).

In T cells, NF- κ B can be activated by several pathways of signal transduction. The engagement of the TCR by major histocompatibility complex (MHC) plus antigen initiates downstream CD3 immunotyrosine activation motif (ITAM) phosphorylation by the Src family kinases, FYN and leukocyte C-terminal src kinase (LCK). Phosphorylated CD3 activates the T cell specific tyrosine kinase, zeta-chain associated protein kinase (ZAP-70), which ultimately trigger calcium release and protein kinase (PK)C activation, respectively. Activation of a specific PKC isoform, PKC μ , connects the above described TCR proximal signaling events to distal events that ultimately lead to NF- κ B activation. Importantly, PKC μ activation is also driven by engagement of the T cell co-stimulatory receptor CD28 by B7 ligands on antigen presenting cells (APCs). In addition, the stimulation of T cells by IL-1 activates NF- κ B as already described before. Once in the nucleus, NF- κ B governs the transcription of numerous genes involved in T cell survival, proliferation, and effector functions (Paul and Schaefer, 2013).

Evidence Supporting this KER

Mice lacking NF- κ B p50 are unable to effectively clear *L. monocytogenes* and are more susceptible to infection with *S. pneumoniae* (Sha et al., 1995).

Biological Plausibility

Although CD4 T cells are able to commit to Th1, Th2 and Th17 lineages in the absence of IL-1R signaling at steady state, these committed CD4 T cells are unable to effectively secrete their cytokines upon TCR ligation. Namely, IL-1 is indispensable for CD4 T cell effector function. (Lin et al, 2015)

RelB deficient mice had an impaired cellular immunity, as observed in contact sensitivity reaction (Weih et al., 1995).

Delayed-type hypersensitivity (DTH) responses were significantly suppressed in IL-1 β -deficient and IL-1 α/β -deficient mice. Lymph node cells derived from antigen-sensitized IL-1 β -deficient and IL-1 α/β -deficient mice and IL-1R type I-deficient mice, exhibited reduced proliferative responses against antigen. (Nambu et al., 2006).

Empirical Evidence

RelB deficient mice had an impaired cellular immunity, as observed in contact sensitivity reaction (Weih et al., 1995).

Quite a few NF- κ B inhibitors have been reported. MG132, bortezomib, curcumin, DHMEQ(Dehydroxymethylepoxyquinomicin), naringin, sorafenib, genistein and parthenolide are some of representatives (Pordanjani and Hosseinimehr, 2016).

Interferon- γ (IFN- γ) production in response to CMV-infected fibroblasts was reduced under the influence of MG132, a proteasome inhibitor as well as a NF- κ B inhibitor, in a dose-dependent manner. A marked reduction was observed at 0.5 μ M. Likewise, CMV-specific cytotoxicity of CD8(+) T cells was decreased in the presence of MG132 (Wang et al., 2011).

In vivo MG132 administration to NC/Nga mice with DNFB-induced dermatitis reduced Th17 cells but maintained the level of Th1 cells, resulting in the alleviation of dermatitis lesions by decreasing both serum IgE hyperproduction and mast cell migration (Ohkusu-Tsukada et al., 2018).

Proteasome inhibitor, bortezomib, potently inhibits the growth of adult T-cell leukemia cells both in vivo and in vitro (Satou et al., 2004). Bortezomib inhibits T-cell function versus infective antigenic stimuli in a dose-dependent manner in vitro (Orciuolo et al., 2007).

DHMEQ, a novel nuclear factor-kappaB inhibitor, induces selective depletion of alloreactive or phytohaemagglutinin-stimulated peripheral blood mononuclear cells, decreases production of T helper type 1 cytokines, and blocks maturation of dendritic cells (Nishioka et al., 2008).

Regarding the suppression of NF- κ B by impaired IL-1 signaling, it was reported that delayed-type hypersensitivity (DTH) responses were significantly suppressed in IL-1 β -deficient and IL-1 α/β -deficient mice. Lymph node cells derived from antigen-sensitized IL-1 β -deficient and IL-1 α/β -deficient mice and IL-1R type I-deficient mice, exhibited reduced proliferative responses against antigen. These data suggest that IL-1 β is necessary for the efficient priming of T cells. In addition, CD4+ T cell-derived IL-1 plays an important role in the activation of DCs during the elicitation phase, resulting in the production of TNF, that activate allergen-specific T cells (Nambu et al., 2006).

Quantitative Understanding of the Linkage

A representative NF- κ B inhibitor, MG132 that suppresses NF- κ B activity at more than 10 nM (Fiedler et al. 1998) suppresses IL-2-

induced activation of STAT5 at 50 mM. (Yu and Malek., 2001).

A representative NF- κ B inhibitor, DHMEQ (1mg/mL) blocked PHA-induced nuclear translocation of NF- κ B in Jurkat cells via inhibition of degradation of I κ B α . Preincubation of peripheral blood mononuclear cells with DHMEQ (1 mg/ml, 3 hr) greatly reduced PHA-stimulated expression of IFN- γ , IL-2 and TNF- α genes (Nishioka et al., 2008).

Response-response relationship

Interferon- γ (IFN- γ) production in response to CMV-infected fibroblasts was reduced under the influence of MG132 in a dose-dependent manner. A marked reduction was observed at 0.5 μ M. Likewise, CMV-specific cytotoxicity of CD8(+) T cells was decreased in the presence of MG132 (Wang et al., 2011).

Bortezomib inhibits T-cell function versus infective antigenic stimuli in a dose-dependent manner in vitro (Orciuolo et al., 2007).

References

- Fiedler, M.A., Wernke-Dollries, K., Stark, J.M. (1998), Inhibition of TNF- α -induced NF- κ B activation and IL-8 release in A549 cells with the proteasome inhibitor MG-132. *Am J Respir Cell Mol Biol* 19: 259-268, 10.1165/ajrcmb.19.2.3149
- Lin, D., Lei, L., Zhang, Y., et al. (2015), Secreted IL-1 α promotes T-cell activation and expansion of CD11b(+) Gr1(+) cells in carbon tetrachloride-induced liver injury in mice. *Eur J Immunol* 45: 2084-2098, 10.1002/eji.201445195
- Nambu, A., Nakae, S., Iwakura, Y. (2006), IL-1 β , but not IL-1 α , is required for antigen-specific T cell activation and the induction of local inflammation in the delayed-type hypersensitivity responses. *Int Immunol* 18: 701-712, 10.1093/intimm/dx1007
- Nishioka, C., Ikezoe, T., Jing, Y., et al. (2008), DHMEQ, a novel nuclear factor- κ B inhibitor, induces selective depletion of alloreactive or phytohemagglutinin-stimulated peripheral blood mononuclear cells, decreases production of T helper type 1 cytokines, and blocks maturation of dendritic cells. *Immunology* 124: 198-205, 10.1111/j.1365-2567.2007.02755.x
- Ohkusu-Tsukada, K., Ito, D., Takahashi, K. (2018), The Role of Proteasome Inhibitor MG132 in 2,4-Dinitrofluorobenzene-Induced Atopic Dermatitis in NC/Nga Mice. *Int Arch Allergy Immunol* 176: 91-100, 10.1159/000488155
- Orciuolo, E., Galimberti, S., Petrini, M. (2007), Bortezomib inhibits T-cell function versus infective antigenic stimuli in a dose-dependent manner in vitro. *Leuk Res* 31: 1026-1027, 10.1016/j.leukres.2006.09.002
- Reinhard, K., Huber, M., Lohoff, M., et al. (2012), The role of NF- κ B activation during protection against Leishmania infection. *Int J Med Microbiol* 302: 230-235, 10.1016/j.ijmm.2012.07.006
- Sha, W.C., Liou, H.C., Tuomanen, E.I., et al. (1995), Targeted disruption of the p50 subunit of NF- κ B leads to multifocal defects in immune responses. *Cell* 80: 321-330,
- van Delft, M.A., Huitema, L.F., Tas, S.W. (2015), The contribution of NF- κ B signalling to immune regulation and tolerance. *Eur J Clin Invest* 45: 529-539, 10.1111/eci.12430
- Wang, Y., Sun, B., Volk, H.D., et al. (2011), Comparative study of the influence of proteasome inhibitor MG132 and ganciclovir on the cytomegalovirus-specific CD8(+) T-cell immune response. *Viral Immunol* 24: 455-461, 10.1089/vim.2011.0038
- Weih, F., Carrasco, D., Durham, S.K., et al. (1995), Multiorgan inflammation and hematopoietic abnormalities in mice with a targeted disruption of RelB, a member of the NF- κ B/Rel family. *Cell* 80: 331-340,
- Yu, A., Malek, T.R. (2001), The proteasome regulates receptor-mediated endocytosis of interleukin-2. *J Biol Chem* 276: 381-385, 10.1074/jbc.M007991200

[Relationship: 2004: Suppression of T cell activation leads to Increase, Increased susceptibility to infection](#)

AOPs Referencing Relationship

AOP Name	Adjacency	Weight of Evidence	Quantitative Understanding
Impaired IL-1R1 signaling leading to increased susceptibility to infection	adjacent	High	High

Evidence Supporting Applicability of this Relationship

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
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Common Term	Scientific Term	Evidence	Links
Homo sapiens	Homo sapiens	High	NCBI
Mus musculus	Mus musculus	High	NCBI
Rattus norvegicus	Rattus norvegicus	High	NCBI

Life Stage Applicability

Life Stage Evidence

All life stages High

Sex Applicability

Sex Evidence

Unspecific High

Key Event Relationship Description

Normal T cell and B cell function is indispensable for host defense mechanism.

Evidence Supporting this KER

The experiments using knockout mice revealed that the lack of IL-1 signaling led to bacterial, tuberculosis or viral infection (Guler et al., 2011; Horino et al., 2009; Juffermans et al., 2000; Tian et al., 2017; Yamada et al., 2000).

Biological Plausibility

To protect the infection from different pathogens, different types of immune response depending on the pathogens are required (reviewed by Soares et al. (Soares et al., 2017)).

1). Type 1 immunity drives resistance to viruses and intracellular bacteria, such as *Listeria monocytogenes*, *Salmonella* spp. and *Mycobacteria* spp., as well as to intracellular protozoan parasites such as *Leishmania* spp. The T helper 1 (T_H1) signature cytokine interferon- γ (IFN γ) has a central role in triggering cytotoxic mechanisms including macrophage polarization towards an antimicrobial response associated with the production of high levels of reactive oxygen species (ROS) and reactive nitrogen species (RNS), activation of CD8⁺ cytotoxic T lymphocytes (CTLs) and natural killer (NK) cells to kill infected cells via the perforin and/or granzyme B-dependent lytic pathway or via the ligation of surface death receptors; and B cell activation towards the production of cytolytic antibodies that target infected cells for complement and Fc receptor-mediated cellular cytotoxicity.

2) Resistance to extracellular metazoan parasites and other large parasites is mediated and/or involves type 2 immunity. Pathogen neutralization is achieved via different mechanisms controlled by T_H2 signature cytokines, including interleukin-4 (IL-4), IL-5 and IL-13, and by additional type 2 cytokines such as thymic stromal lymphopoietin (TSLP), IL-25 or IL-33, secreted by damaged cell. T_H2 signature cytokines drive B cell activation towards the production of high-affinity pathogen-specific IgG1 and IgE antibodies that function via Fc-dependent mechanisms to trigger the activation of eosinophils, mast cells and basophils, expelling pathogens across epithelia.

3) T_H17 immunity confers resistance to extracellular bacteria such as *Klebsiella pneumoniae*, *Escherichia coli*, *Citrobacter rodentium*, *Bordetella pertussis*, *Porphyromonas gingivalis* and *Streptococcus pneumoniae*, and also to fungi such as *Candida albicans*, *Coccidioides posadasii*, *Histoplasma capsulatum* and *Blastomyces dermatitidis*. Activation of T_H17 cells by cognate T cell receptor (TCR–MHC class II interactions and activation of group 3 innate lymphoid cells (ILC3s) via engagement of IL-1 receptor (IL-1R) by IL-1 β secreted from damaged cells lead to the recruitment and activation of neutrophils. T_H17 immunopathology is driven to a large extent by products of neutrophil activation, such as ROS and elastase.

Based on these evidences, the insufficient T cell or B cell function causes impaired resistance to infection.

Empirical Evidence

Administration of IL-1R antagonist or neutralizing antibody such as IL-1Ra (generic anakinra), canakinumab (anti-IL-1 β antibody) and rilonacept (soluble IL-1R) led to the suppression of downstream phenomena, which included internalization of IL-1 (Dripps et al., 1991), production of PGE₂ (Hannum et al., 1990; Seckinger et al., 1990), IL-6 (Goh et al., 2014), and T cell proliferation (Seckinger et al., 1990).

Since these inhibitors became available to treat some of autoimmune syndromes, it became clear that these inhibitors increased the frequency of serious bacterial infection (De Benedetti et al., 2018; Genovese et al., 2004; Imagawa et al., 2013; Kullenberg et al., 2016; Lachmann et al., 2009; Lequerre et al., 2008; Migkos et al., 2015; Schlesinger et al., 2012; Yokota et al., 2017).

Certain pharmaceutical agents known as calcineurin inhibitors that suppress T cell function and are commonly used to prevent organ rejection of transplant recipients or to treat autoimmune disorders, also have an immunosuppressive side effect, known to

lead to an increase in the following opportunistic infections: fungal/yeast (e.g., *Cryptococcus neoformans*); viral (esp. herpes-family viruses such as Epstein-Barr virus [EBV], cytomegalovirus [CMV]); atypical mycobacterial (e.g., mycoplasma, *Nocardia*, *Listeria*, mycobacteria); and parasitic (e.g., toxoplasmosis) infections (reviewed by Singh (2005)).

Quantitative Understanding of the Linkage

Luster et al (1993) demonstrated that Concanavalin A response of splenocytes showed the linear dose-response relationship with the host resistance to *Listeria monocytogenes* or *Streptococcus pneumoniae*.

Response-response relationship

Luster et al (1993) demonstrated that Concanavalin A response of splenocytes showed the linear dose-response relationship with the host resistance to *Listeria monocytogenes* or *Streptococcus pneumoniae*.

References

- De Benedetti, F., Gattorno, M., Anton, J., et al. (2018), Canakinumab for the Treatment of Autoinflammatory Recurrent Fever Syndromes. *N Engl J Med* 378: 1908-1919, 10.1056/NEJMoa1706314
- Dripps, D.J., Brandhuber, B.J., Thompson, R.C., et al. (1991), Interleukin-1 (IL-1) receptor antagonist binds to the 80-kDa IL-1 receptor but does not initiate IL-1 signal transduction. *J Biol Chem* 266: 10331-10336,
- Fleischmann, R.M., Schechtman, J., Bennett, R., et al. (2003), Anakinra, a recombinant human interleukin-1 receptor antagonist (r-metHuIL-1ra), in patients with rheumatoid arthritis: A large, international, multicenter, placebo-controlled trial. *Arthritis Rheum* 48: 927-934, 10.1002/art.10870
- Genovese, M.C., Cohen, S., Moreland, L., et al. (2004), Combination therapy with etanercept and anakinra in the treatment of patients with rheumatoid arthritis who have been treated unsuccessfully with methotrexate. *Arthritis Rheum* 50: 1412-1419, 10.1002/art.20221
- Goh, A.X., Bertin-Maghit, S., Ping Yeo, S., et al. (2014), A novel human anti-interleukin-1beta neutralizing monoclonal antibody showing in vivo efficacy. *MAbs* 6: 765-773, 10.4161/mabs.28614
- Guler, R., Parihar, S.P., Spohn, G., et al. (2011), Blocking IL-1alpha but not IL-1beta increases susceptibility to chronic Mycobacterium tuberculosis infection in mice. *Vaccine* 29: 1339-1346, 10.1016/j.vaccine.2010.10.045
- Hannum, C.H., Wilcox, C.J., Arend, W.P., et al. (1990), Interleukin-1 receptor antagonist activity of a human interleukin-1 inhibitor. *Nature* 343: 336-340, 10.1038/343336a0
- Horino, T., Matsumoto, T., Ishikawa, H., et al. (2009), Interleukin-1 deficiency in combination with macrophage depletion increases susceptibility to *Pseudomonas aeruginosa* bacteremia. *Microbiol Immunol* 53: 502-511, 10.1111/j.1348-0421.2009.00143.x
- Imagawa, T., Nishikomori, R., Takada, H., et al. (2013), Safety and efficacy of canakinumab in Japanese patients with phenotypes of cryopyrin-associated periodic syndrome as established in the first open-label, phase-3 pivotal study (24-week results). *Clin Exp Rheumatol* 31: 302-309,
- Juffermans, N.P., Florquin, S., Camoglio, L., et al. (2000), Interleukin-1 signaling is essential for host defense during murine pulmonary tuberculosis. *J Infect Dis* 182: 902-908, 10.1086/315771
- Kullenberg, T., Lofqvist, M., Leinonen, M., et al. (2016), Long-term safety profile of anakinra in patients with severe cryopyrin-associated periodic syndromes. *Rheumatology (Oxford)* 55: 1499-1506, 10.1093/rheumatology/kew208
- Lachmann, H.J., Kone-Paut, I., Kuemmerle-Deschner, J.B., et al. (2009), Use of canakinumab in the cryopyrin-associated periodic syndrome. *N Engl J Med* 360: 2416-2425, 10.1056/NEJMoa0810787
- Lequerre, T., Quartier, P., Rosellini, D., et al. (2008), Interleukin-1 receptor antagonist (anakinra) treatment in patients with systemic-onset juvenile idiopathic arthritis or adult onset Still disease: preliminary experience in France. *Ann Rheum Dis* 67: 302-308, 10.1136/ard.2007.076034
- Luster, M.I., Portier, C., Pait, D.G., et al. (1993), Risk assessment in immunotoxicology. II. Relationships between immune and host resistance tests. *Fundam Appl Toxicol* 21: 71-82, 10.1006/faat.1993.1074
- Migkos, M.P., Somarakis, G.A., Markatseli, T.E., et al. (2015), Tuberculous pyomyositis in a rheumatoid arthritis patient treated with anakinra. *Clin Exp Rheumatol* 33: 734-736,
- Schlesinger, N., Alten, R.E., Bardin, T., et al. (2012), Canakinumab for acute gouty arthritis in patients with limited treatment options: results from two randomised, multicentre, active-controlled, double-blind trials and their initial extensions. *Ann Rheum Dis* 71: 1839-1848, 10.1136/annrheumdis-2011-200908
- Seckinger, P., Kaufmann, M.T., Dayer, J.M. (1990), An interleukin 1 inhibitor affects both cell-associated interleukin 1-induced T cell proliferation and PGE2/collagenase production by human dermal fibroblasts and synovial cells. *Immunobiology* 180: 316-327,

10.1016/s0171-2985(11)80295-0

Singh, N. (2005), Infectious complications in organ transplant recipients with the use of calcineurin-inhibitor agent-based immunosuppressive regimens. *Curr Opin Infect Dis* 18: 342-345, 10.1097/01.qco.0000172698.52408.be

Soares, M.P., Teixeira, L., Moita, L.F. (2017), Disease tolerance and immunity in host protection against infection. *Nat Rev Immunol* 17: 83-96, 10.1038/nri.2016.136

Tian, T., Jin, M.Q., Dubin, K. (2017), IL-1R Type 1-Deficient Mice Demonstrate an Impaired Host Immune Response against Cutaneous Vaccinia Virus Infection. 198: 4341-4351, 10.4049/jimmunol.1500106

Yamada, H., Mizumo, S., Horai, R., et al. (2000), Protective role of interleukin-1 in mycobacterial infection in IL-1 alpha/beta double-knockout mice. *Lab Invest* 80: 759-767,

Yokota, S., Imagawa, T., Nishikomori, R., et al. (2017), Long-term safety and efficacy of canakinumab in cryopyrin-associated periodic syndrome: results from an open-label, phase III pivotal study in Japanese patients. *Clin Exp Rheumatol* 35 Suppl 108: 19-26,