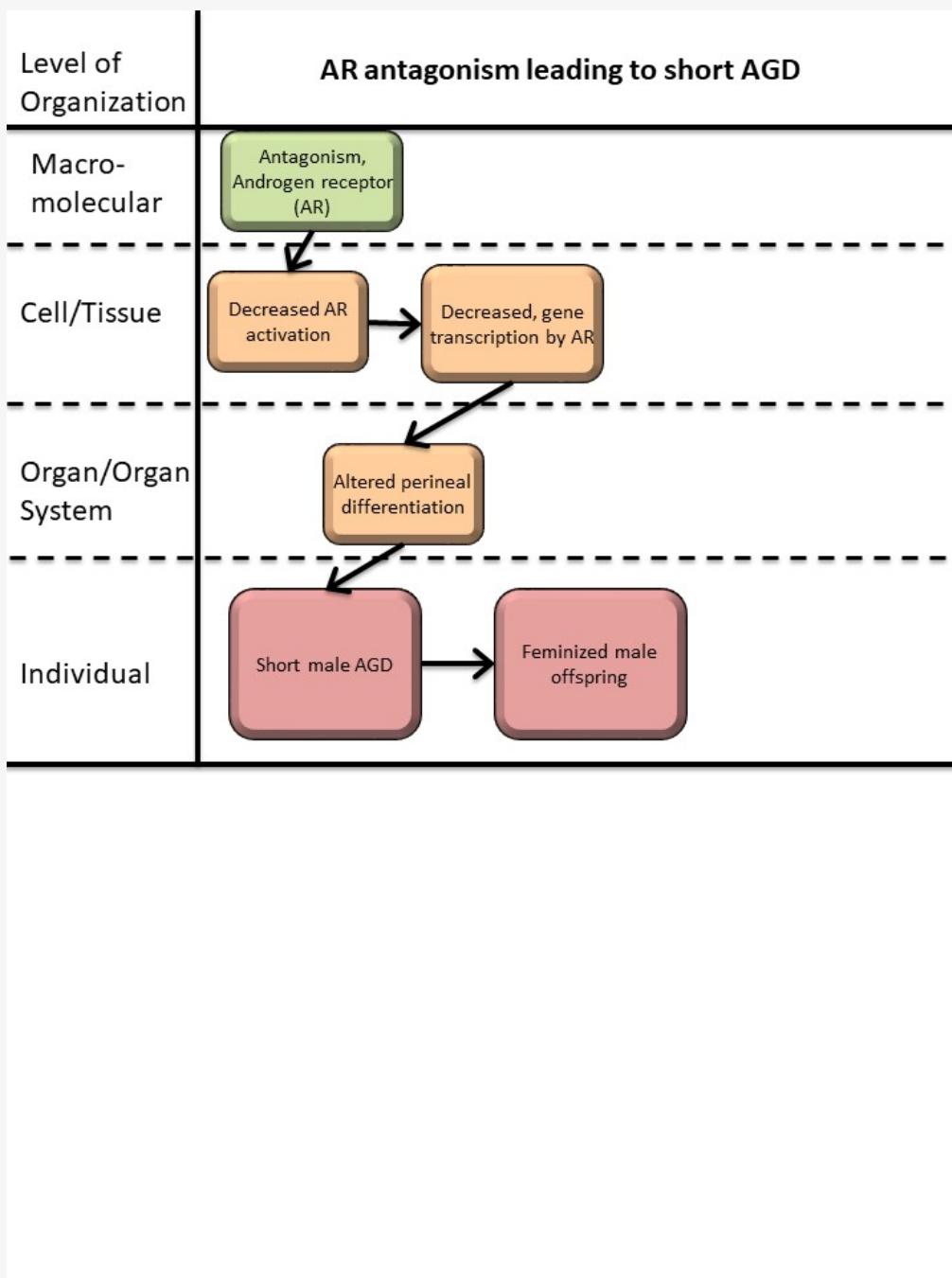


AOP ID and Title:

AOP 306: Androgen receptor (AR) antagonism leading to short anogenital distance (AGD) in male (mammalian) offspring
Short Title: AR antagonism leading to short AGD

Graphical Representation**Authors**

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Status

Author status	OECD status	OECD project	SAAOP status
Under development: Not open for comment. Do not cite	Under Development	1.90	Included in OECD Work Plan

	Author status	OECD status	OECD project	SAAOP status
Abstract				
This AOP links Androgen receptor antagonism during fetal life with short anogenital distance (AGD) in male offspring. A short AGD around birth is a marker for feminization of male fetuses and is associated with male reproductive disorders, including reduced fertility in adulthood. Although a short AGD is not necessarily 'adverse' from a human health perspective, it is considered an 'adverse outcome' in OECD test guidelines; AGD measurements are mandatory in specific tests for developmental and reproductive toxicity in chemical risk assessment (TG 443, TG 421/422, TG 414).				
The AR is a nuclear receptor involved in the transcriptional regulation of various target genes during development and adulthood across species. Its main ligand is testosterone and dihydrotestosterone (DHT). Under normal physiological conditions, testosterone produced mainly by the testicles, is converted in peripheral tissues by 5 α -reductase into DHT, which in turn binds AR and activates downstream target genes. AR signaling is necessary for normal masculinization of the developing fetus, including differentiation of the levator ani/bulbocavernosus (LABC) muscle complex in male fetuses. The LABC complex does not develop in the absence, or low levels of, androgen signaling, as in female fetuses.				
The key events in this pathway is antagonism of the AR in target cells of the primitive perineal region, which leads to inactivation of the AR and failure to properly masculinize the perineum/LABC complex. In this instance, the local levels of testosterone or DHT may be normal, but prevented from binding the AR.				
Summary of the AOP				
Events				
Molecular Initiating Events (MIE), Key Events (KE), Adverse Outcomes (AO)				
Sequence	Type	Event ID	Title	Short name
MIE	26	Antagonism, Androgen receptor	Antagonism, Androgen receptor	
KE	1614	Decrease, androgen receptor activation	Decrease, AR activation	
KE	1687	decrease, transcription of genes by AR	decrease, transcription of genes by AR	
AO	1688	anogenital distance (AGD), decreased	AGD, decreased	
Key Event Relationships				
Upstream Event	Relationship Type	Downstream Event	Evidence	Quantitative Understanding
Antagonism, Androgen receptor	adjacent	Decrease, androgen receptor activation	High	High
Decrease, androgen receptor activation	adjacent	decrease, transcription of genes by AR	High	Moderate
decrease, transcription of genes by AR	adjacent	anogenital distance (AGD), decreased	Moderate	Low
Antagonism, Androgen receptor	non-adjacent	anogenital distance (AGD), decreased	Moderate	Low
Decrease, androgen receptor activation	non-adjacent	anogenital distance (AGD), decreased		
Stressors				
Name	Evidence			
Finasteride	High			
Flutamide	High			
Finasteride				

Intrauterine exposure in rats can result in shorter male AGD in male offspring as reported in:

Bowman et al (2003), *Toxicol Sci* 74:393-406; doi: 10.1093/toxsci/kfg128

Christiansen et al (2009), *Environ Health Perspect* 117:1839-1846; doi: 10.1289/ehp.0900689

Schwartz et al (2019), *Toxicol Sci* 169:303-311; doi: 10.1093/toxsci/kfz046

Flutamide

Finasteride is a selective androgen receptor (AR) antagonist (Simard et al 1986) that has been shown to induce shorter male AGD in rats after in utero exposure (Foster & Harris 2005; Hass et al 2007; Kita et al 2016; McIntyre et al 2001; Mylchreest et al 1999; Scott et al 2007; Welsh et al 2007).

References:

Foster & Harris (2005), *Toxicol Sci* 85:1024-1032; doi: 10.1093/toxsci/kfi159

Hass et al (2007), *Environ Health Perspect* 115(suppl 1):122-128; doi: 10.1289/ehp.0360

Kita et al (2016), *Toxicology* 368-369:152-161; doi: 10.1016/j.tox.2016.08.021

McIntyre et al (2001), *Toxicol Sci* 62:236-249; doi: 10.1093/toxsci/62.2.236

Mylchreest et al (1999), *Toxicol Appl Pharmacol* 156:81-95; doi: 10.1006/taap.1999.8643

Scott et al (2007), *Endocrinology* 148:2027-2036; doi: 10.1210/en.2006-1622

Simard et al (1986), *Mol Cell Endocrinol* 44:261-270; doi: 10.1016/0303-7207(86)90132-2

Overall Assessment of the AOP

Domain of Applicability

Life Stage Applicability

Life Stage Evidence

Pregnancy High

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
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human	Homo sapiens	Moderate	NCBI
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rat	Rattus norvegicus	High	NCBI
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mouse	Mus musculus	Moderate	NCBI
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Sex Applicability

Sex Evidence

Male High

References

1. Schwartz CL, Christiansen S, Vinggaard AM, Axelstad M, Hass U and Svingen T (2019), Anogenital distance as a toxicological or clinical marker for fetal androgen action and risk for reproductive disorders. *Arch Toxicol* 93: 253-272.

Appendix 1

List of MIEs in this AOP

[Event: 26: Antagonism, Androgen receptor](#)

Short Name: Antagonism, Androgen receptor

AOPs Including This Key Event

AOP ID and Name	Event Type
Aop:306 - Androgen receptor (AR) antagonism leading to short anogenital distance (AGD) in male (mammalian) offspring	MolecularInitiatingEvent
Aop:344 - Androgen receptor (AR) antagonism leading to nipple retention (NR) in male (mammalian) offspring	MolecularInitiatingEvent
Aop:345 - Androgen receptor (AR) antagonism leading to decreased fertility in females	MolecularInitiatingEvent
Aop:372 - Androgen receptor antagonism leading to testicular cancer	MolecularInitiatingEvent
Aop:477 - Androgen receptor (AR) antagonism leading to hypospadias in male offspring	MolecularInitiatingEvent
Aop:476 - Adverse Outcome Pathways diagram related to PBDEs associated male reproductive toxicity	MolecularInitiatingEvent

Stressors

Name
Mercaptobenzole
Triticonazole
Flusilazole
Epoxiconazole
Prochloraz
Propiconazole
Tebuconazole
Flutamide
Cyproterone acetate
Vinclozolin

Biological Context

Level of Biological Organization

Molecular

Cell term

Cell term

eukaryotic cell

Evidence for Perturbation by Stressor

Overview for Molecular Initiating Event

A large number of drugs and chemicals have been shown to antagonise the AR using various AR reporter gene assays. The AR is specifically targeted in AR-sensitive cancers, for example the use of the anti-androgenic drug flutamide in treating prostate cancer ([Alapi & Fischer, 2006](#)). Flutamide has also been used in several rodent *in vivo* studies showing anti-androgenic effects (feminization of male offspring) evident by e.g. short anogenital distance (AGD) in males ([Foster & Harris, 2005](#); [Hass et al, 2007](#); [Kita et al, 2016](#)). QSAR models can predict AR antagonism for a wide range of chemicals, many of which have shown *in vitro* antagonistic potential ([Vinggaard et al, 2008](#)).

Triticonazole

Using hAR-EcoScreen Assay, triticonazole showed a LOEC for antagonisms of 0.2 uM and an IC50 of 0.3 (± 0.01) uM ([Draskau et al. 2019](#))

Flusilazole

Using hAR-EcoScreen Assay, flusilazole showed a LOEC for antagonisms of 0.8 uM and an IC50 of 2.8 (± 0.1) uM ([Draskau et al. 2019](#))►

Epoxiconazole

Using transiently AR-transfected CHO cells, epoxiconazole showed a LOEC of 1.6 uM and an IC50 of 10 uM ([Kjærstad et al. 2010](#))

Prochloraz

Using transiently AR-transfected CHO cells, prochloraz showed a LOEC of 6.3 uM and an IC50 of 13 uM ([Kjærstad et al. 2010](#))

Propiconazole

Using transiently AR-transfected CHO cells, propiconazole showed a LOEC of 12.5 uM and an IC50 of 18 uM ([Kjærstad et al. 2010](#))

Tebuconazole

Using transiently AR-transfected CHO cells, tebuconazole showed a LOEC of 3.1 uM and an IC50 of 8.1 uM ([Kjærstad et al. 2010](#))

Flutamide

Using the AR-CALUX reporter assay in antagonism mode, flutamide showed an IC50 of 1.3 uM ([Sonneveld et al. 2005](#)).

Cyproterone acetate

Using the AR-CALUX reporter assay in antagonism mode, cyproterone acetate showed an IC50 of 7.1 nM ([Sonneveld et al. 2005](#)).

Vinclozolin

Using the AR-CALUX reporter assay in antagonism mode, vinclozolin showed an IC50 of 1.0 uM ([Sonneveld et al. 2005](#)).

Domain of Applicability

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
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Vertebrates	Vertebrates	High	NCBI
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Life Stage Applicability

Life Stage	Evidence
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During development and at adulthood	High
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Sex Applicability

Sex	Evidence
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Mixed	High
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Both the DNA-binding and ligand-binding domains of the AR are highly evolutionary conserved, whereas the transactivation domain show more divergence which may affect AR-mediated gene regulation across species ([Davey & Grossmann, 2016](#)). Despite certain inter-species differences, AR function mediated through gene expression is highly conserved, with mutations studies from both humans and rodents showing strong correlation for AR-dependent development and function ([Walters et al. 2010](#)). Likewise in fish, androgens are important for development of sexual characteristics (Ogino et al., 2014, 2023). One difference that must be mentioned is that in teleost fish, 11-ketotestosterone is the main androgen in addition to testosterone and DHT and that most teleosts have two *ar* orthologs, *ara* and *arb*, with *arb* functioning in a similar manner to the AR in other vertebrates (Ogino et al., 2023).

This KE is applicable for both sexes, across developmental stages into adulthood, in numerous cells and tissues and across

vertebrate taxa

Key Event Description

The androgen receptor (AR) and its function

The AR is a ligand-activated transcription factor belonging to the steroid hormone nuclear receptor family ([Davey & Grossmann, 2016](#)). The AR has three domains: the N-terminal domain, the DNA-binding domain and the ligand-binding domain, with the latter being most evolutionary conserved. Testosterone (T) and the more biologically active dihydrotestosterone (DHT) are endogenous ligands for the AR ([MacLean et al. 1993](#); [MacLeod et al. 2010](#); [Schwartz et al. 2019](#)). In teleost fishes, 11-ketotestosterone is the second main ligand ([Schuppe et al. 2020](#)). Human AR mutations and mouse knock-out models have established a pivotal role for the AR in masculinization and spermatogenesis ([Walters et al. 2010](#)). Apart from the essential role for AR in male reproductive development and function ([Walters et al. 2010](#)), the AR is also expressed in many other tissues and organs such as bone, muscles, ovaries, and the immune system ([Rana et al. 2014](#)).

AR antagonism as Key Event

The main function of the AR is to activate gene transcription in cells. Canonical signaling occurs by ligands (androgens) binding to AR in the cytoplasm which results in translocation to the cell nucleus, receptor dimerization and binding to specific regulatory DNA sequences ([Heemers & Tindall, 2007](#)). The gene targets regulated by AR activation depends on cell/tissue type and what stage of development activation occur, and is, for instance, dependent on available co-factors. Apart from the canonical signaling pathway, AR can also initiate cytoplasmic signaling pathways with other functions than the nuclear pathway, for instance rapid change in cell function by ion transport changes ([Heinlein & Chang, 2002](#)) and association with Src kinase to activate MAPK/ERK signaling and activation of the PI3K/Akt pathway ([Leung & Sadar, 2017](#)).

How it is Measured or Detected

AR antagonism can be measured in vitro by transient or stable transactivation assays to evaluate nuclear receptor activation. There is already a validated assay for AR (ant)agonism adopted by the OECD, Test No. 458: *Stably Transfected Human Androgen Receptor Transcriptional Activation Assay for Detection of Androgenic Agonist and Antagonist Activity of Chemicals* ([OECD, 2016](#)). The stably transfected AR-EcoScreenTM cells ([Satoh et al. 2004](#)) should be used for the assay and are freely available from the Japanese Collection of Research Bioresources (JCRB) Cell Bank under reference number JCRB1328.

Other assays include the AR-CALUX reporter gene assay that is derived from human U2-OS cells stably transfected with the human AR and an AR responsive reporter gene ([Sonneveld et al. 2004](#); [van der Burg et al. 2010](#)), various transiently transfected reporter cell lines ([Körner et al. 2004](#)), and more.

The recently developed AR dimerization assay provides an assay with an improved ability to measure potential stressor-mediated disruption of dimerization/activation ([Lee et al. 2021](#)).

The Rapid Androgen Disruption Activity Reporter (RADAR) assay included in OECD test guideline no. 251 detects AR antagonism in vivo in fish ([OECD 2022](#)).

References

Davey RA, Grossmann M (2016) Androgen Receptor Structure, Function and Biology: From Bench to Bedside. *Clin Biochem Rev* **37**: 3-15

Heemers HV, Tindall DJ (2007) Androgen receptor (AR) coregulators: a diversity of functions converging on and regulating the AR transcriptional complex. *Endocr Rev* **28**: 778-808

Heinlein CA, Chang C (2002) The roles of androgen receptors and androgen-binding proteins in nongenomic androgen actions. *Mol Endocrinol* **16**: 2181-2187

Körner W, Vinggaard AM, Térouanne B, Ma R, Wieloch C, Schlumpf M, Sultan C, Soto AM (2004) Interlaboratory comparison of four in vitro assays for assessing androgenic and antiandrogenic activity of environmental chemicals. *Environ Health Perspect* **112**: 695-702

Lee SH, Hong KY, Seo H, Lee HS, Park Y (2021) Mechanistic insight into human androgen receptor-mediated endocrine-disrupting potentials by a stable bioluminescence resonance energy transfer-based dimerization assay. *Chem Biol Interact* **349**: 109655

Leung, J. K., & Sadar, M. D. (2017). Non-Genomic Actions of the Androgen Receptor in Prostate Cancer. *Frontiers in Endocrinology*, **8**. <https://doi.org/10.3389/fendo.2017.00002>

MacLean HE, Chu S, Warne GL, Zajac JD (1993) Related individuals with different androgen receptor gene deletions. *J Clin Invest* **91**: 1123-1128

MacLeod DJ, Sharpe RM, Welsh M, Fiskin M, Scott HM, Hutchison GR, Drake AJ, van den Driesche S (2010) Androgen action in the masculinization programming window and development of male reproductive organs. *Int J Androl* **33**: 279-287

OECD. (2016) Test No. 458: Stably Transfected Human Androgen Receptor Transcriptional Activation Assay for Detection of Androgenic Agonist and Antagonist Activity of Chemicals. *OECD Guidelines for the Testing of Chemicals, Section 4*, Paris.

OECD (2022). Test No. 251: [Rapid Androgen Disruption Activity Reporter \(RADAR\) assay](#). Paris: OECD Publishing doi:10.1787/da264d82-en.

Ogino, Y., Ansai, S., Watanabe, E., Yasugi, M., Katayama, Y., Sakamoto, H., et al. (2023). Evolutionary differentiation of androgen receptor is responsible for sexual characteristic development in a teleost fish. *Nat. Commun.* 2023 **14** 14, 1–16. doi:10.1038/s41467-023-37026-6.

Ogino, Y., Hirakawa, I., Inohaya, K., Sumiya, E., Miyagawa, S., Denslow, N., et al. (2014). Bmp7 and Lef1 Are the Downstream Effectors of Androgen Signaling in Androgen-Induced Sex Characteristics Development in Medaka. *Endocrinology* 155, 449–462. doi:10.1210/EN.2013-1507.

Rana K, davey RA, Zajac JD (2014) Human androgen deficiency: insights gained from androgen receptor knockout mouse models. *Asian J Androl* **16**: 169-177

Satoh K, Ohyama K, Aoki N, Iida M, Nagai F (2004) Study on anti-androgenic effects of bisphenol a diglycidyl ether (BADGE), bisphenol F diglycidyl ether (BFDGE) and their derivatives using cells stably transfected with human androgen receptor, AR-EcoScreen. *Food Chem Toxicol* **42**: 983-993

Schuppe, E. R., Miles, M. C., and Fuxjager, M. J. (2020). Evolution of the androgen receptor: Perspectives from human health to dancing birds. *Mol. Cell. Endocrinol.* 499, 110577. doi:10.1016/J.MCE.2019.110577

Schwartz CL, Christiansen S, Vinggaard AM, Axelstad M, Hass U, Svingen T (2019) Anogenital distance as a toxicological or clinical marker for fetal androgen action and risk for reproductive disorders. *Arch Toxicol* **93**: 253-272

Sonneveld E, Jansen HJ, Riteco JA, Brouwer A, van der Burg B (2005) Development of androgen- and estrogen-responsive bioassays, members of a panel of human cell line-based highly selective steroid-responsive bioassays. *Toxicol Sci* **83**: 136-148

van der Burg B, Winter R, Man HY, Vangenechten C, Berckmans P, Weimer M, Witters H, van der Linden S (2010) Optimization and prevalidation of the in vitro AR CALUX method to test androgenic and antiandrogenic activity of compounds. *Reprod Toxicol* **30**: 18-24

Walters KA, Simanainen U, Handelsman DJ (2010) Molecular insights into androgen actions in male and female reproductive function from androgen receptor knockout models. *Hum Reprod Update* **16**: 543-558

List of Key Events in the AOP

[Event: 1614: Decrease, androgen receptor activation](#)

Short Name: Decrease, AR activation

AOPs Including This Key Event

AOP ID and Name	Event Type
Aop:288 - Inhibition of 17α-hydroxylase/C 10,20-lyase (Cyp17A1) activity leads to birth reproductive defects (cryptorchidism) in male (mammals)	KeyEvent
Aop:305 - 5α-reductase inhibition leading to short anogenital distance (AGD) in male (mammalian) offspring	KeyEvent
Aop:306 - Androgen receptor (AR) antagonism leading to short anogenital distance (AGD) in male (mammalian) offspring	KeyEvent
Aop:307 - Decreased testosterone synthesis leading to short anogenital distance (AGD) in male (mammalian) offspring	KeyEvent
Aop:344 - Androgen receptor (AR) antagonism leading to nipple retention (NR) in male (mammalian) offspring	KeyEvent
Aop:372 - Androgen receptor antagonism leading to testicular cancer	KeyEvent
Aop:477 - Androgen receptor (AR) antagonism leading to hypospadias in male offspring	KeyEvent

Biological Context

Level of Biological Organization

Tissue

Domain of Applicability

Taxonomic Applicability

Term Scientific Term Evidence Links

Vertebrates	Vertebrates	High	NCBI
Term	Scientific Term	Evidence	Links
Life Stage Applicability			

Life Stage	Evidence
During development and at adulthood	High

Sex Applicability

Sex	Evidence
Mixed	High

This KE is considered broadly applicable across vertebrate taxa as all vertebrate animals express the AR in numerous cells and tissues where it regulates gene transcription required for developmental processes and functions.

Key Event Description

This KE refers to decreased activation of the androgen receptor (AR) as occurring in complex biological systems such as tissues and organs *in vivo*. It is thus considered distinct from KEs describing either blocking of AR or decreased androgen synthesis.

The AR is a nuclear transcription factor with canonical AR activation regulated by the binding of the androgens such as testosterone or dihydrotestosterone (DHT). Thus, AR activity can be decreased by reduced levels of steroidal ligands (testosterone, DHT) or the presence of compounds interfering with ligand binding to the receptor (Davey & Grossmann, 2016; Gao et al., 2005).

In the inactive state, AR is sequestered in the cytoplasm of cells by molecular chaperones. In the classical (genomic) AR signaling pathway, AR activation causes dissociation of the chaperones, AR dimerization and translocation to the nucleus to modulate gene expression. AR binds to the androgen response element (Davey & Grossmann, 2016; Gao et al., 2005). AR does not, however, act alone in regulating gene transcription, but together with other co-factors that may differ between cells and tissues and life stages. In this way, the functional consequence of AR activation is cell- and tissue-dependent.

Ligand-bound AR may also associate with cytoplasmic and membrane-bound proteins to initiate cytoplasmic signaling pathways with other functions than the nuclear pathway. Non-genomic AR signaling includes association with Src kinase to activate MAPK/ERK signaling and activation of the PI3K/Akt pathway. Decreased AR activity may therefore be a decrease in the genomic and/or non-genomic AR signaling pathways (Leung & Sadar, 2017).

How it is Measured or Detected

This KE specifically focuses on decreased *in vivo* activation, with most methods that can be used to measure AR activity carried out *in vitro*. They provide indirect information about the KE and are described in lower tier MIE/KEs (see MIE/KE-26 for AR antagonism, KE-1690 for decreased T levels and KE-1613 for decreased dihydrotestosterone levels). In this way, this KE is a placeholder for tissue-specific responses to AR activation or inactivation that will depend on the adverse outcome (AO) for which it is included.

It should be mentioned that the Rapid Androgen Disruption Activity Reporter (RADAR) assay included in OECD test guideline no. 251 detects AR antagonism *in vivo* in fish (OECD 2022).

References

Davey, R. A., & Grossmann, M. (2016). Androgen Receptor Structure, Function and Biology: From Bench to Bedside. *The Clinical Biochemist Reviews*, 37(1), 3–15.

Gao, W., Bohl, C. E., & Dalton, J. T. (2005). Chemistry and structural biology of androgen receptor. *Chemical Reviews*, 105(9), 3352–3370. <https://doi.org/10.1021/cr020456u>

Hutson, J. M. (1985). A biphasic model for the hormonal control of testicular descent. *The Lancet*, 24, 419–421. [https://doi.org/https://doi.org/10.1016/S0140-6736\(85\)92739-4](https://doi.org/https://doi.org/10.1016/S0140-6736(85)92739-4)

Kaftanovskaya, E. M., Huang, Z., Barbara, A. M., de Gendt, K., Verhoeven, G., Gorlov, I. P., & Agoulnik, A. I. (2012). Cryptorchidism in mice with an androgen receptor ablation in the gubernaculum testis. *Molecular Endocrinology*, 26(4), 598–607. <https://doi.org/10.1210/me.2011-1283>

Lee, S. H., Hong, K. Y., Seo, H., Lee, H. S., & Park, Y. (2021). Mechanistic insight into human androgen receptor-mediated endocrine-disrupting potentials by a stable bioluminescence resonance energy transfer-based dimerization assay. *Chemico-Biological Interactions*, 349. <https://doi.org/10.1016/j.cbi.2021.109655>

Leung, J. K., & Sadar, M. D. (2017). Non-Genomic Actions of the Androgen Receptor in Prostate Cancer. *Frontiers in Endocrinology*, 8. <https://doi.org/10.3389/fendo.2017.00002>

OECD (2022). Test No. 251: Rapid Androgen Disruption Activity Reporter (RADAR) assay. Paris: OECD Publishing doi:10.1787/da264d82-en.

Pang, T. P. S., Clarke, M. v., Ghasem-Zadeh, A., Lee, N. K. L., Davey, R. A., & MacLean, H. E. (2012). A physiological role for androgen actions in the absence of androgen receptor DNA binding activity. *Molecular and Cellular Endocrinology*, 348(1), 189–197. <https://doi.org/10.1016/j.mce.2011.08.017>

[Event: 1687: decrease, transcription of genes by AR](#)**Short Name:** decrease, transcription of genes by AR**AOPs Including This Key Event**

AOP ID and Name	Event Type
Aop:306 - Androgen receptor (AR) antagonism leading to short anogenital distance (AGD) in male (mammalian) offspring	KeyEvent
Aop:372 - Androgen receptor antagonism leading to testicular cancer	KeyEvent

Biological Context**Level of Biological Organization**

Cellular

List of Adverse Outcomes in this AOP[Event: 1688: anogenital distance \(AGD\), decreased](#)**Short Name:** AGD, decreased**Key Event Component**

Process	Object	Action
androgen receptor signaling pathway	Musculature of male perineum	disrupted

AOPs Including This Key Event

AOP ID and Name	Event Type
Aop:305 - 5α-reductase inhibition leading to short anogenital distance (AGD) in male (mammalian) offspring	AdverseOutcome
Aop:306 - Androgen receptor (AR) antagonism leading to short anogenital distance (AGD) in male (mammalian) offspring	AdverseOutcome
Aop:307 - Decreased testosterone synthesis leading to short anogenital distance (AGD) in male (mammalian) offspring	AdverseOutcome
Aop:476 - Adverse Outcome Pathways diagram related to PBDEs associated male reproductive toxicity	AdverseOutcome

Stressors

Name
Butylparaben
p,p'-DDE
Bis(2-ethylhexyl) phthalate
Dexamethasone
Fenitrothion
Finasteride
Flutamide

Ketoconazole**Name**

Linuron

Prochloraz

Procymidone

Triticonazole

Vinclozolin

di-n-hexyl phthalate

Dicyclohexyl phthalate

butyl benzyl phthalate

monobenzyl phthalate

di-n-heptyl phthalate

Biological Context**Level of Biological Organization**

Tissue

Organ term**Organ term**

perineum

Evidence for Perturbation by Stressor**Butylparaben**

Butylparaben has been shown to cause decreased male AGD in rats following intrauterine exposure to 500 and 1000 mg/kg bw/day ([Boberg et al. 2016](#); [Zhang et al. 2014](#)). A separate study using 600 mg/kg bw/day did not see an effect on male AGD ([Boberg et al. 2008](#)).

p,p'-DDE

p,p'DDE has been shown to cause decreased male AGD in rats following intrauterine exposure to 100-200 mg/kg bw/day ([Loeffler & Peterson. 1999](#); [Wolf et al. 1999](#)).

Bis(2-ethylhexyl) phthalate

DEHP has been shown to cause decreased male AGD in rats following intrauterine exposure to 300-1500 mg/kg bw/day ([Christiansen et al. 2010](#); [Gray et al. 2000](#); [Howdeshell et al. 2007](#); [Jarfelt et al. 2005](#); [Kita et al. 2016](#); [Li et al. 2013](#); [Lin et al. 2009](#); [Moore et al. 2001](#); [Nardelli et al. 2017](#); [Saillenfait et al. 2009](#); [Wolf et al. 1999](#)).

Dexamethasone

Dexamethasone has been shown to cause decreased male AGD in rats following intrauterine exposure to 0.1 mg/kg bw/day ([Van den Driesche et al. 2012](#)).

Fenitrothion

Fenitrothion has been shown to cause decreased male AGD in rats following intrauterine exposure to 25 mg/kg bw/day ([Turner et al. 2002](#)).

Finasteride

Finasteride has been shown to cause decreased male AGD in rats following intrauterine exposure to 100 mg/kg bw/day ([Bowman et al. 2003](#)).

Flutamide

Flutamide has been shown to cause decreased male AGD in rats following intrauterine exposure to doses between 16-100 mg/kg bw/day ([Foster & Harris, 2005](#); [Hass et al. 2007](#); [Kita et al. 2016](#); [McIntyre et al. 2001](#); [Mylchreest et al. 1999](#); [Scott et al. 2007](#); [Welsh et al. 2007](#)).

Ketoconazole

Ketoconazole has been shown to cause decreased male AGD in rats following intrauterine exposure to 50 mg/kg bw/day in one study ([Taxvig et al. 2008](#)), but no effect in another study using same dose ([Wolf et al. 1999](#)).

Linuron

Linuron has been shown to cause decreased male AGD in rats following intrauterine exposure to 50-100 mg/kg bw/day ([Hotchkiss et al. 2004](#); [McIntyre et al. 2002](#); [Wolf et al. 1999](#)).

Prochloraz

Prochloraz has been shown to cause decreased male AGD in rats following intrauterine exposure to 150-250 mg/kg bw/day ([Laier et al. 2006](#); [Noriega et al. 2005](#)).

Procymidone

Procymidone has been shown to cause decreased male AGD in rats following intrauterine exposure to doses between 50-150 mg/kg bw/day ([Hass et al. 2012](#); [Hass et al. 2007](#); [Wolf et al. 1999](#)).

Triticonazole

Triticonazole has been shown to cause decreased male AGD in rats following intrauterine exposure to 150 and 450 mg/kg bw/day ([Draskau et al. 2019](#)).

Vinclozolin

Vinclozolin has been shown to cause decreased male AGD in rats following intrauterine exposure to doses between 50-200 mg/kg bw/day ([Christiansen et al. 2009](#); [Gray et al. 1994](#); [Hass et al. 2007](#); [Matsuura et al. 2005](#); [Ostby et al. 1999](#); [Schneider et al. 2011](#); [Wolf et al. 2004](#)).

di-n-hexyl phthalate

DnHP has been shown to cause decreased male AGD in rats following intrauterine exposure to 500-750 mg/kg bw/day ([Saillenfait et al. 2009a](#); [Saillenfait et al. 2009b](#)).

Dicyclohexyl phthalate

DCHP has been shown to cause decreased male AGD in rats following intrauterine exposure to 350-750 mg/kg bw/day ([Aydoğan Ahbab & Barlas, 2015](#); [Hoshino et al. 2005](#); [Saillenfait et al. 2009a](#)).

butyl benzyl phthalate

BBP has been shown to cause decreased male AGD in rats following intrauterine exposure to 500-1000 mg/kg bw/day ([Ema & Miyawaki, 2002](#); [Gray et al. 2000](#); [Hotchkiss et al. 2004](#); [Nagao et al. 2000](#); [Tyl et al. 2004](#)).

monobenzyl phthalate

MBeP has been shown to cause decreased male AGD in rats following intrauterine exposure to 375 mg/kg bw/day ([Ema et al. 2003](#)).

di-n-heptyl phthalate

DHPP has been shown to cause decreased male AGD in rats following intrauterine exposure to 1000 mg/kg bw/day ([Saillenfait et al. 2011](#)).

Domain of Applicability

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
human	Homo sapiens	Moderate	NCBI
rat	Rattus norvegicus	High	NCBI
mouse	Mus musculus	High	NCBI

Life Stage Applicability

Life Stage Evidence

Foetal High

Sex Applicability

Sex Evidence

Male High

A short AGD in male offspring is a marker of insufficient androgen action during critical fetal developmental stages ([Schwartz et al. 2019](#); [Welsh et al. 2008](#)). A short AGD is thus a sign of undervirilization, which is also associated with a series of male reproductive disorders, including genital malformations and infertility in humans ([Juul et al. 2014](#); [Skakkebaek et al. 2001](#)).

There are numerous human epidemiological studies showing associations with intrauterine exposure to anti-androgenic chemicals and short AGD in newborn boys alongside other reproductive disorders ([Schwartz et al. 2019](#)). This underscores the human relevance of this AO. However, in reproductive toxicity studies and chemical risk assessment, rodents (rats and mice) are what is tested on. The list of chemicals inducing short male AGD in male rat offspring is extensive, as evidenced by the 'stressor' list and reviewed by ([Schwartz et al. 2019](#)).

Key Event Description

The anogenital distance (AGD) refers to the distance between anus and the external genitalia. In rodents and humans, the male AGD is approximately twice the length as the female AGD ([Salazar-Martinez et al. 2004](#); [Schwartz et al. 2019](#)). This sexual dimorphism is a consequence of sex hormone-dependent development of secondary sexual characteristics ([Schwartz et al. 2019](#)). In males, it is believed that androgens (primarily DHT) activate AR-positive cells in non-myotitic cells in the fetal perineum region to initiate differentiation of the perineal *levator ani* and *bulbocavernosus* (LABC) muscle complex ([Ipulan et al. 2014](#)). This AR-dependent process occurs within a critical window of development, around gestational days 15-18 in rats ([MacLeod et al. 2010](#)). In females, the absence of DHT prevents this masculinization effect from occurring.

The involvement of androgens in masculinization of the male fetus, including the perineum, has been known for a very long time ([Jost, 1953](#)), and AGD has historically been used to, for instance, sex newborn kittens. It is now well established that the AGD in newborns is a proxy readout for the intrauterine sex hormone milieu the fetus was developing. Too low androgen levels in XY fetuses makes the male AGD shorter, whereas excess (ectopic) androgen levels in XX fetuses makes the female AGD longer, in humans and rodents ([Schwartz et al. 2019](#)).

How it is Measured or Detected

The AGD is a morphometric measurement carried out by trained technicians (rodents) or medical staff (humans).

In rodent studies AGD is assessed as the distance between the genital papilla and the anus, and measured using a stereomicroscope with a micrometer eyepiece. The AGD index (AGDi) is often calculated by dividing AGD by the cube root of the body weight. It is important in statistical analysis to use litter as the statistical unit. This is done when more than one pup from each litter is examined. Statistical analyses are adjusted using litter as an independent, random and nested factor. AGD are analysed using body weight as covariate as recommended in Guidance Document 151 ([OECD, 2013](#)).

Regulatory Significance of the AO

In regulatory toxicology, the AGD is mandatory inclusions in OECD test guidelines used to test for developmental and reproductive toxicity of chemicals. Guidelines include 'TG 443 extended one-generation study', 'TG 421/422 reproductive toxicity screening studies' and 'TG 414 developmental toxicity study'.

References

[Aydoğan Ahbab M, Barlas N \(2015\) Influence of in utero di-n-hexyl phthalate and dicyclohexyl phthalate on fetal testicular](#)

development in rats. *Toxicol Lett* **233**: 125-137

Boberg J, Axelstad M, Svingen T, Mandrup K, Christiansen S, Vinggaard AM, Hass U (2016) Multiple endocrine disrupting effects in rats perinatally exposed to butylparaben. *Toxicol Sci* **152**: 244-256

Boberg J, Metzdorff S, Wortziger R, Axelstad M, Brokken L, Vinggaard AM, Dalgaard M, Nelleman C (2008) Impact of diisobutyl phthalate and other PPAR agonists on steroidogenesis and plasma insulin and leptin levels in fetal rats. *Toxicology* **250**: 75-81

Bowman CJ, Barlow NJ, Turner KJ, Wallace DG, Foster PM (2003) Effects of in utero exposure to finasteride on androgen-dependent reproductive development in the male rat. *Toxicol Sci* **74**: 393-406

Christiansen S, Boberg J, Axelstad M, Dalgaard M, Vinggaard AM, Metzdorff SB, Hass U (2010) Low-dose perinatal exposure to di(2-ethylhexyl) phthalate induces anti-androgenic effects in male rats. *Reprod Toxicol* **30**: 313-321

Christiansen S, Scholze M, Dalgaard M, Vinggaard AM, Axelstad M, Kortenkamp A, Hass U (2009) Synergistic disruption of external male sex organ development by a mixture of four antiandrogens. *Environ Health Perspect* **117**: 1839-1846

Draskau MK, Boberg J, Taxvig C, Pedersen M, Frandsen HL, Christiansen S, Svingen T (2019) In vitro and in vivo endocrine disrupting effects of the azole fungicides triticonazole and flusilazole. *Environ Pollut* **255**: 113309

Ema M, Miyawaki E (2002) Effects on development of the reproductive system in male offspring of rats given butyl benzyl phthalate during late pregnancy. *Reprod Toxicol* **16**: 71-76

Ema M, Miyawaki E, Hirose A, Kamata E (2003) Decreased anogenital distance and increased incidence of undescended testes in fetuses of rats given monobenzyl phthalate, a major metabolite of butyl benzyl phthalate. *Reprod Toxicol* **17**: 407-412

Foster PM, Harris MW (2005) Changes in androgen-mediated reproductive development in male rat offspring following exposure to a single oral dose of flutamide at different gestational ages. *Toxicol Sci* **85**: 1024-1032

Gray LE, Jr., Ostby J, Furr J, Price M, Veeramachaneni DN, Parks L (2000) Perinatal exposure to the phthalates DEHP, BBP, and DINP, but not DEP, DMP, or DOTP, alters sexual differentiation of the male rat. *Toxicol Sci* **58**: 350-365

Gray LEJ, Ostby JS, Kelce WR (1994) Developmental effects of an environmental antiandrogen: the fungicide vinclozolin alters sex differentiation of the male rat. *Toxicol Appl Pharmacol* **129**: 46-52

Hass U, Boberg J, Christiansen S, Jacobsen PR, Vinggaard AM, Taxvig C, Poulsen ME, Herrmann SS, Jensen BH, Petersen A, Clemmensen LH, Axelstad M (2012) Adverse effects on sexual development in rat offspring after low dose exposure to a mixture of endocrine disrupting pesticides. *Reprod Toxicol* **34**: 261-274

Hass U, Scholze M, Christiansen S, Dalgaard M, Vinggaard AM, Axelstad M, Metzdorff SB, Kortenkamp A (2007) Combined exposure to anti-androgens exacerbates disruption of sexual differentiation in the rat. *Environ Health Perspect* **115 Suppl. 1**: 122-128

Hoshino N, Iwai M, Okazaki Y (2005) A two-generation reproductive toxicity study of dicyclohexyl phthalate in rats. *J Toxicol Sci* **30 Spec No**: 79-96

Hotchkiss AK, Parks-Salduti LG, Ostby JS, Lambright C, Furr J, Vandenbergh JG, Gray LEJ (2004) A mixture of the "antiandrogens" linuron and butyl benzyl phthalate alters sexual differentiation of the male rat in a cumulative fashion. *Biol Reprod* **71**: 1852-1861

Howdeshell KL, Furr J, Lambright CR, Rider CV, Wilson VS, Gray LE, Jr. (2007) Cumulative effects of dibutyl phthalate and diethylhexyl phthalate on male rat reproductive tract development: altered fetal steroid hormones and genes. *Toxicol Sci* **99**: 190-202

Ipulan LA, Suzuki K, Sakamoto Y, Murashima A, Imai Y, Omori A, Nakagata N, Nishinakamura R, Valasek P, Yamada G (2014) Nonmyocytic androgen receptor regulates the sexually dimorphic development of the embryonic bulbocavernosus muscle. *Endocrinology* **155**: 2467-2479

Jarfelt K, Dalgaard M, Hass U, Borch J, Jacobsen H, Ladefoged O (2005) Antiandrogenic effects in male rats perinatally exposed to a mixture of di(2-ethylhexyl) phthalate and di(2-ethylhexyl) adipate. *Reprod Toxicol* **19**: 505-515

Jost A (1953) Problems of fetal endocrinology: The gonadal and hypophyseal hormones. *Recent Prog Horm Res* **8**: 379-418

Juul A, Almstrup K, Andersson AM, Jensen TK, Jorgensen N, Main KM, Rajpert-De Meyts E, Toppari J, Skakkebaek NE (2014) Possible fetal determinants of male infertility. *Nat Rev Endocrinol* **10**: 553-562

Kita DH, Meyer KB, Venturelli AC, Adams R, Machado DL, Morais RN, Swan SH, Gennings C, Martino-Andrade AJ (2016) Manipulation of pre and postnatal androgen environments and anogenital distance in rats. *Toxicology* **368-369**: 152-161

Laier P, Metzdorff SB, Borch J, Hagen ML, Hass U, Christiansen S, Axelstad M, Kledal T, Dalgaard M, McKinnell C, Brokken LJ, Vinggaard AM (2006) Mechanisms of action underlying the antiandrogenic effects of the fungicide prochloraz. *Toxicol Appl Pharmacol* **213**: 2

Li M, Qiu L, Zhang Y, Hua Y, Tu S, He Y, Wen S, Wang Q, Wei G (2013) Dose-related effect by maternal exposure to di-(2-ethylhexyl) phthalate plasticizer on inducing hypospadiac male rats. *Environ Toxicol Pharmacol* **35**: 55-60

Lin H, Lian QQ, Hu GX, Jin Y, Zhang Y, Hardy DO, Chen GR, Lu ZQ, Sottas CM, Hardy MP, Ge RS (2009) In utero and lactational exposures to diethylhexyl-phthalate affect two populations of Leydig cells in male Long-Evans rats. *Biol Reprod* **80**: 882-888

Loeffler IK, Peterson RE (1999) Interactive effects of TCDD and p,p'-DDE on male reproductive tract development in in utero and lactationally exposed rats. *Toxicol Appl Pharmacol* **154**: 28-39

MacLeod DJ, Sharpe RM, Welsh M, Fiskin M, Scott HM, Hutchison GR, Drake AJ, van den Driesche S (2010) Androgen action in the masculinization programming window and development of male reproductive organs. *Int J Androl* **33**: 279-287

Matsuura I, Saitoh T, Ashina M, Wako Y, Iwata H, Toyota N, Ishizuka Y, Namiki M, Hoshino N, Tsuchitani M (2005) Evaluation of a two-generation reproduction toxicity study adding endpoints to detect endocrine disrupting activity using vinclozolin. *J Toxicol Sci* **30 Spec No**: 163-168

McIntyre BS, Barlow NJ, Foster PM (2001) Androgen-mediated development in male rat offspring exposed to flutamide in utero: permanence and correlation of early postnatal changes in anogenital distance and nipple retention with malformations in androgen-dependent tissues. *Toxicol Sci* **62**: 236-249

McIntyre BS, Barlow NJ, Sar M, Wallace DG, Foster PM (2002) Effects of in utero linuron exposure on rat Wolffian duct development. *Reprod Toxicol* **16**: 131-139

Melching-Kollmuss S, Fussell KC, Schneider S, Buesen R, Groeters S, Strauss V, van Ravenzwaay B (2017) Comparing effect levels of regulatory studies with endpoints derived in targeted anti-androgenic studies: example prochloraz. *Arch Toxicol* **91**: 143-162

Moore RW, Rudy TA, Lin TM, Ko K, Peterson RE (2001) Abnormalities of sexual development in male rats with in utero and lactational exposure to the antiandrogenic plasticizer Di(2-ethylhexyl) phthalate. *Environ Health Perspect* **109**: 229-237

Mylchreest E, Sar M, Cattley RC, Foster PM (1999) Disruption of androgen-regulated male reproductive development by di(n-butyl) phthalate during late gestation in rats is different from flutamide. *Toxicol Appl Pharmacol* **156**: 81-95

Nagao T, Ohta R, Marumo H, Shindo T, Yoshimura S, Ono H (2000) Effect of butyl benzyl phthalate in Sprague-Dawley rats after gavage administration: a two-generation reproductive study. *Reprod Toxicol* **14**: 513-532

Nardelli TC, Albert O, Lalancette C, Culty M, Hales BF, Robaire B (2017) In utero and lactational exposure study in rats to identify replacements for di(2-ethylhexyl) phthalate. *Sci Rep* **7**: 3862

Noriega NC, Ostby J, Lambright C, Wilson VS, Gray LE, Jr. (2005) Late gestational exposure to the fungicide prochloraz delays the onset of parturition and causes reproductive malformations in male but not female rat offspring. *Biol Reprod* **72**: 1324-1335

OECD. (2013) Guidance document in support of the test guideline on the extended one generation reproductive toxicity study No. 151.

Ostby J, Kelce WR, Lambright C, Wolf CJ, Mann P, Gray CLJ (1999) The fungicide procymidone alters sexual differentiation in the male rat by acting as an androgen-receptor antagonist in vivo and in vitro. *Toxicol Ind Health* **15**: 80-93

Saillenfait AM, Gallissot F, Sabaté JP (2009a) Differential developmental toxicities of di-n-hexyl phthalate and dicyclohexyl phthalate administered orally to rats. *J Appl Toxicol* **29**: 510-521

Saillenfait AM, Roudot AC, Gallissot F, Sabaté JP (2011) Prenatal developmental toxicity studies on di-n-heptyl and di-n-octyl phthalates in Sprague-Dawley rats. *Reprod Toxicol* **32**: 268-276

Saillenfait AM, Sabaté JP, Gallissot F (2009b) Effects of in utero exposure to di-n-hexyl phthalate on the reproductive development of the male rat. *Reprod Toxicol* **28**: 468-476

Salazar-Martinez E, Romano-Riquer P, Yanez-Marquez E, Longnecker MP, Hernandez-Avila M (2004) Anogenital distance in human male and female newborns: a descriptive, cross-sectional study. *Environ Health* **3**: 8

Schneider S, Kaufmann W, Strauss V, van Ravenzwaay B (2011) Vinclozolin: a feasibility and sensitivity study of the ILSI-HESI F1-extended one-generation rat reproduction protocol. *Regulatory Toxicology and Pharmacology* **59**: 91-100

Schwartz CL, Christiansen S, Vinggaard AM, Axelstad M, Hass U, Svingen T (2019) Anogenital distance as a toxicological or clinical marker for fetal androgen action and risk for reproductive disorders. *Arch Toxicol* **93**: 253-272

Scott HM, Hutchison GR, Mahood IK, Hallmark N, Welsh M, De Gendt K, Verhoeven H, O'Shaughnessy P, Sharpe RM (2007) Role of androgens in fetal testis development and dysgenesis. *Endocrinology* **148**: 2027-2036

Skakkebaek NE, Rajpert-De Meyts E, Main KM (2001) Testicular dysgenesis syndrome: an increasingly common developmental disorder with environmental aspects. *Hum Reprod* **16**: 972-978

Taxvig C, Vinggaard AM, Hass U, Axelstad M, Metzdorff S, Nellemann C (2008) Endocrine-disrupting properties in vivo of widely used azole fungicides. *Int J Androl* **31**: 170-177

Turner KJ, Barlow NJ, Struve MF, Wallace DG, Gaido KW, Dorman DC, Foster PM (2002) Effects of in utero exposure to the organophosphate insecticide fenitrothion on androgen-dependent reproductive development in the Crl:CD(SD)BR rat. *Toxicol Sci* **68**: 174-183

Tyl RW, Myers CB, Marr MC, Fail PA, Seely JC, Brine DR, Barter RA, Butala JH (2004) Reproductive toxicity evaluation of dietary butyl benzyl phthalate (BBP) in rats. *Reprod Toxicol* **18**: 241-264

Van den Driesche S, Kolovos P, Platts S, Drake AJ, Sharpe RM (2012) Inter-relationship between testicular dysgenesis and Leydig cell function in the masculinization programming window in the rat. *PLoS one* **7**: e30111

Welsh M, Saunders PT, Fiskin M, Scott HM, Hutchison GR, Smith LB, Sharpe RM (2008) Identification in rats of a programming window for reproductive tract masculinization, disruption of which leads to hypospadias and cryptorchidism. *J Clin Invest* **118**: 1479-1490

Welsh M, Saunders PT, Sharpe RM (2007) The critical time window for androgen-dependent development of the Wolffian duct in the rat. *Endocrinology* **148**: 3185-3195

Wolf CJ, LeBlanc GA, Gray LE, Jr. (2004) Interactive effects of vinclozolin and testosterone propionate on pregnancy and sexual differentiation of the male and female SD rat. *Toxicol Sci* **78**: 135-143

Wolf CJ, Lambright C, Mann P, Price M, Cooper RL, Ostby J, Gray CLJ (1999) Administration of potentially antiandrogenic pesticides (procymidone, linuron, iprodione, chlozolinate, p,p'-DDE, and ketoconazole) and toxic substances (dibutyl- and diethylhexyl phthalate, PCB 169, and ethane dimethane sulphonate) during sexual differentiation produces diverse profiles of reproductive malformations in the male rat. *Toxicol Ind Health* **15**: 94-118

Zhang L, Dong L, Ding S, Qiao P, Wang C, Zhang M, Zhang L, Du Q, Li Y, Tang N, Chang B (2014) Effects of n-butylparaben on steroidogenesis and spermatogenesis through changed E₂ levels in male rat offspring. *Environ Toxicol Pharmacol* **37**: 705-717

Appendix 2

List of Key Event Relationships in the AOP

List of Adjacent Key Event Relationships

[Relationship: 2130: Antagonism, Androgen receptor leads to Decrease, AR activation](#)

AOPs Referencing Relationship

AOP Name	Adjacency	Weight of Evidence	Quantitative Understanding
Androgen receptor (AR) antagonism leading to short anogenital distance (AGD) in male (mammalian) offspring	adjacent	High	High
Androgen receptor (AR) antagonism leading to nipple retention (NR) in male (mammalian) offspring	adjacent	High	
Androgen receptor (AR) antagonism leading to hypospadias in male offspring	adjacent	High	

Evidence Supporting Applicability of this Relationship

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
Vertebrates	Vertebrates	High	NCBI

Life Stage Applicability

Life Stage	Evidence
During development and at adulthood	High

Sex Applicability

Sex	Evidence
Mixed	High

The AR is expressed throughout vertebrate taxa and its DNA and ligand binding domains are highly conserved (Davey & Grossmann, 2016). AR activity is important for sexual development and reproduction in both males and females (Prizant et al., 2014; Walters et al., 2010). AR function is required during development, puberty and adulthood.

Key Event Relationship Description

The androgen receptor (AR) is a ligand-activated steroid hormone nuclear receptor (Davey & Grossmann, 2016). In its inactive state, the AR locates to the cytoplasm (Roy et al., 2001). When activated, the AR translocates to the nucleus, dimerizes, and, together with co-regulators, binds to specific DNA regulatory sequences to regulate gene transcription (Davey & Grossmann, 2016) (Lamont and Tindall, 2010). This is considered the canonical AR signaling pathway. The AR can also activate non-genomic signalling (Jin et al., 2013). However, this KER focuses on the canonical pathway.

The two main AR ligands are the androgens testosterone (T) and the more potent dihydrotestosterone (DHT), whereas another main androgen in teleost fishes is 11-ketotestosterone (Schuppe et al., 2020). Androgens bind to the AR to mediate downstream androgenic responses, such as male development and masculinization (Rey, 2021; Walters et al., 2010). Antagonism of the AR would decrease AR activation and therefore the downstream AR-mediated effects.

Evidence Supporting this KER

Biological Plausibility

The biological plausibility for this KER is considered high.

The AR belongs to the steroid hormone nuclear receptor family. The AR has 3 main domains essential for its activity, the N-terminal domain, the ligand binding domain, and the DNA binding domain (Roy et al., 2001). Ligands, such as T and DHT, must bind to the ligand binding domain to activate AR allowing it to fulfill its role as a transcription factor. The binding of the ligand induces a change in AR conformation allowing it to translocate to the nucleus and congregate into a subnuclear compartment (Marcelli et al., 2006; Roy et al., 2001) homodimerize and bind to the DNA target sequences and regulate transcription of target genes. Regulation of AR target genes is greatly facilitated by numerous co-factors. Active AR signaling is essential for male reproduction and sexual development and is also crucial in several other tissues and organs such as ovaries, the immune system, bones, and muscles (Ogino et al., 2011; Prizant et al., 2014; Rey, 2021; William H. Walker, 2021).

AR antagonists can compete with or prevent in different ways AR ligand binding, thereby preventing AR activation. Antagonism of the AR can prevent translocation to the nucleus, compartmentalization, dimerization and DNA binding. Consequently, AR cannot regulate transcription of target genes and androgen signalling is disrupted. This can be observed using different AR activation assays such as AR dimerization, translocation, DNA binding or transcriptional activity assays (Brown et al., 2023; OECD, 2020).

Empirical Evidence

The empirical evidence for this KER is considered high

The effects of AR antagonism have been shown in many studies *in vivo* and *in vitro*.

Several stressors can act as antagonists of the AR and lead to decreased AR activation. Some of these are detailed in an AOP key event relationship report by (Pedersen et al., 2022) and shown below, exhibiting evidence of dose-concordance:

Stressors

- Cyproterone acetate: Using the AR-CALUX reporter assay in antagonism mode, cyproterone acetate showed an IC50 of 7.1 nM (Sonneveld, 2005)
- Epoxiconazole: Using transiently AR-transfected CHO cells, epoxiconazole showed a LOEC of 1.6 μ M and an IC50 of 10 μ M (Kjærstad et al., 2010).
- Flutamide: Using the AR-CALUX reporter assay in antagonism mode, flutamide showed an IC50 of 1.3 μ M (Sonneveld, 2005).
- Flusilazole: Using hAR-EcoScreen Assay, triticonazole showed a LOEC for antagonisms of 0.8 μ M and an IC50 of 2.8 (\pm 0.1) μ M (Draskau et al., 2019).
- Prochloraz: Using transiently AR-transfected CHO cells, prochloraz showed a LOEC of 6.3 μ M and an IC50 of 13 μ M (Kjærstad et al., 2010).
- Propiconazole: Using transiently AR-transfected CHO cells, propiconazole showed a LOEC of 12.5 μ M and an IC50 of 18 μ M (Kjærstad et al., 2010).
- Tebuconazole: Using transiently AR-transfected CHO cells, tebuconazole showed a LOEC of 3.1 μ M and an IC50 of 8.1 μ M (Kjærstad et al., 2010).
- Triticonazole: Using hAR-EcoScreen Assay, triticonazole showed a LOEC for antagonisms of 0.2 μ M and an IC50 of 0.3 (\pm 0.01) μ M (Draskau et al., 2019).
- Vinclozolin: Using the AR-CALUX reporter assay in antagonism mode, vinclozolin showed an IC50 of 1.0 μ M (Sonneveld, 2005). (Pedersen et al., 2022)

Other evidence:

Known AR antagonists are used for treatment of AR-sensitive cancers such as flutamide for prostate cancer (Mahler et al., 1998).

Quantitative Understanding of the Linkage

Response-response relationship

The quantitative relationship between AR antagonism and AR activation will depend on the type of antagonist.

Time-scale

Nuclear translocation in HeLa cells transfected with AR-GFP show a response within 2 hours after ligand exposure (Marcelli et al., 2006; Szafran et al., 2008). Another assay focusing on AR binding to promoters in LNCaP cells has shown that after ligand binding, AR is able to translocate and bind to the DNA sequences within 15min showing the speed of AR activation (Kang et al., 2002).

Known Feedforward/Feedback loops influencing this KER

AR antagonism can lead to increased AR transcript stability and levels as a compensatory mechanism in prostate cancer cells (Dart et al., 2020). In turn, in presence of increased AR levels, AR antagonists can exhibit agonistic activity (Chen et al., 2003).

References

Brown, E. C., Hallinger, D. R., Simmons, S. O., Puig-Castellví, F., Eilebrecht, E., Arnold, L., & Bioscience, P. A. (2023). High-throughput AR dimerization assay identifies androgen disrupting chemicals and metabolites. *Front. Toxicol.*, 5, 1134783. <https://doi.org/10.3389/ftox.2023.1134783>

Chen, C. D., Welsbie, D. S., Tran, C., Baek, S. H., Chen, R., Vessella, R., Rosenfeld, M. G., & Sawyers, C. L. (2003). A R T I C L E S Molecular determinants of resistance to antiandrogen therapy. *NATURE MEDICINE*, 10(1). <https://doi.org/10.1038/nm972>

Dart, D. A., Ashelford, K., & Jiang, W. G. (2020). *AR mRNA stability is increased with AR-antagonist resistance via 3'UTR variants*. <https://doi.org/10.1530/EC-19-0340>

Davey, R. A., & Grossmann, M. (2016). Androgen Receptor Structure, Function and Biology: From Bench to Bedside. In *Androgen Receptor Biology Clin Biochem Rev* (Vol. 37, Issue 1).

Draskau, M. K., Boberg, J., Taxvig, C., Pedersen, M., Frandsen, H. L., Christiansen, S., & Svingen, T. (2019). In vitro and in vivo endocrine disrupting effects of the azole fungicides triticonazole and flusilazole. *Environmental Pollution*, 255, 113309. <https://doi.org/10.1016/j.envpol.2019.113309>

Jin, H. J., Kim, J., & Yu, J. (2013). Androgen receptor genomic regulation. In *Translational Andrology and Urology* (Vol. 2, Issue 3, pp. 158–177). AME Publishing Company. <https://doi.org/10.3978/j.issn.2223-4683.2013.09.01>

Kang, Z., Pirskanen, A., Jänne, O. A., & Palvimo, J. J. (2002). Involvement of Proteasome in the Dynamic Assembly of the Androgen Receptor Transcription Complex. *Journal of Biological Chemistry*, 277(50), 48366–48371. <https://doi.org/10.1074/jbc.M209074200>

Kjærstad, M. B., Taxvig, C., Nellemann, C., Vinggaard, A. M., & Andersen, H. R. (2010). Endocrine disrupting effects in vitro of conazole antifungals used as pesticides and pharmaceuticals. *Reproductive Toxicology*, 30(4), 573–582. <https://doi.org/10.1016/j.reprotox.2010.07.009>

Lamont, K. R., and Tindall, D. J. (2010). Androgen Regulation of Gene Expression. *Adv. Cancer Res.* 107, 137–162. doi:10.1016/S0065-230X(10)07005-3.

Mahler, C., Verhelst, J., and Denis, L. (1998). Clinical pharmacokinetics of the antiandrogens and their efficacy in prostate cancer. *Clin. Pharmacokinet.* 34, 405–417. doi:10.2165/00003088-199834050-00005/METRICS.

Marcelli, M., Stenoien, D. L., Szafran, A. T., Simeoni, S., Agoulnik, I. U., Weigel, N. L., Moran, T., Mikic, I., Price, J. H., & Mancini, M. A. (2006). Quantifying effects of ligands on androgen receptor nuclear translocation, intranuclear dynamics, and solubility. *Journal of Cellular Biochemistry*, 98(4), 770–788. <https://doi.org/10.1002/jcb.20593>

OECD (2020). Test No. 458: Stably Transfected Human Androgen Receptor Transcriptional Activation Assay for Detection of Androgenic Agonist and Antagonist Activity of Chemicals. OECD Guide. Paris: OECD Publishing doi:10.1787/9789264264366-en.

Ogino, Y., Miyagawa, S., Katoh, H., Prins, G. S., Iguchi, T., & Yamada, G. (2011). Essential functions of androgen signaling emerged through the developmental analysis of vertebrate sex characteristics. *Evolution & Development*, 13(3), 315–325. <https://doi.org/10.1111/j.1525-142X.2011.00482.x>

Pedersen, E. B., Christiansen, S., & Svingen, T. (2022). AOP key event relationship report: Linking androgen receptor antagonism with nipple retention. *Current Research in Toxicology*, 3, 100085. <https://doi.org/10.1016/j.crtox.2022.100085>

Prizant, H., Gleicher, N., & Sen, A. (2014). Androgen actions in the ovary: balance is key. *Journal of Endocrinology*, 222(3), R141–R151. <https://doi.org/10.1530/JOE-14-0296>

Rey, R. A. (2021). The Role of Androgen Signaling in Male Sexual Development at Puberty. *Endocrinology*, 162(2). <https://doi.org/10.1210/endocr/bqaa215>

Roy, A. K., Tyagi, R. K., Song, C. S., Lavrovsky, Y., Ahn, S. C., Oh, T. S., & Chatterjee, B. (2001). Androgen receptor: Structural domains and

functional dynamics after ligand-receptor interaction. *Annals of the New York Academy of Sciences*, 949, 44–57. <https://doi.org/10.1111/j.1749-6632.2001.tb04001.x>

Sonneveld, E. (2005). Development of Androgen- and Estrogen-Responsive Bioassays, Members of a Panel of Human Cell Line-Based Highly Selective Steroid-Responsive Bioassays. *Toxicological Sciences*, 83(1), 136–148. <https://doi.org/10.1093/toxsci/kfi005>

Szafran, A. T., Szwarc, M., Marcelli, M., & Mancini, M. A. (2008). Androgen Receptor Functional Analyses by High Throughput Imaging: Determination of Ligand, Cell Cycle, and Mutation-Specific Effects. *PLoS ONE*, 3(11), e3605. <https://doi.org/10.1371/journal.pone.0003605>

Walters, K. A., Simanainen, U., & Handelsman, D. J. (2010). Molecular insights into androgen actions in male and female reproductive function from androgen receptor knockout models. In *Human Reproduction Update* (Vol. 16, Issue 5, pp. 543–558). Hum Reprod Update. <https://doi.org/10.1093/humupd/dmq003>

William H. Walker. (2021). Androgen Actions in the Testis and the Regulation of Spermatogenesis. In *Advances in Experimental Medicine and Biology: Vol. volume 1381* (pp. 175–203).

[Relationship: 2128: Decrease, AR activation leads to decrease, transcription of genes by AR](#)

AOPs Referencing Relationship

AOP Name	Adjacency	Weight of Evidence	Quantitative Understanding
Androgen receptor (AR) antagonism leading to short anogenital distance (AGD) in male (mammalian) offspring	adjacent	High	Moderate

Evidence Supporting Applicability of this Relationship

Taxonomic Applicability

Term	Scientific Term	Evidence	Links
Vertebrates	Vertebrates	High	NCBI

Life Stage Applicability

Life Stage	Evidence
During development and at adulthood	High

Sex Applicability

Sex	Evidence
Mixed	High

[Relationship: 2129: decrease, transcription of genes by AR leads to AGD, decreased](#)

AOPs Referencing Relationship

AOP Name	Adjacency	Weight of Evidence	Quantitative Understanding
Androgen receptor (AR) antagonism leading to short anogenital distance (AGD) in male (mammalian) offspring	adjacent	Moderate	Low

List of Non Adjacent Key Event Relationships

[Relationship: 2123: Antagonism, Androgen receptor leads to AGD, decreased](#)

AOPs Referencing Relationship

AOP Name	Adjacency	Weight of Evidence	Quantitative Understanding
Androgen receptor (AR) antagonism leading to short anogenital distance (AGD) in male (mammalian) offspring	non-adjacent	Moderate	Low

[Relationship: 2820: Decrease, AR activation leads to AGD, decreased](#)

AOPs Referencing Relationship

AOP Name	Adjacency	Weight of Evidence	Quantitative Understanding
<u>5α-reductase inhibition leading to short anogenital distance (AGD) in male (mammalian) offspring</u>	non-adjacent		
<u>Androgen receptor (AR) antagonism leading to short anogenital distance (AGD) in male (mammalian) offspring</u>	non-adjacent		
<u>Decreased testosterone synthesis leading to short anogenital distance (AGD) in male (mammalian) offspring</u>	non-adjacent		